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The prevalence of onchocerciasis in Africa and Yemen, 2000–2018: a geospatial analysis

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1.0 GATHER compliance

Supplementary Table 1. Guidelines for Accurate and Transparent Health Estimates Reporting (GATHER) checklist.

Item #	Checklist item	Reported on page #
Objectiv	es and Funding	
1	Define the indicator(s), populations (including age, sex, and geographic entities), and time period(s) for which estimates were made.	Main Text: Introduction, Methods (Data inputs); Methods (Covariates)
2	List the funding sources for the work.	Main text: Acknowledgments
Data Inp	outs	
For all de	ata inputs from multiple sources that are synthesized as part of the study:	
3	Describe how the data were identified and how the data were accessed.	Main Text: Methods (Data inputs); Supplementary Information: 3.2 Systematic review
4	Specify the inclusion and exclusion criteria. Identify all ad-hoc exclusions.	Main Text: Methods (Data inputs); 3.0 Supplementary data; Supplementary Information: 3.2 Systematic review
5	Provide information on all included data sources and their main characteristics. For each data source used, report reference information or contact name/institution, population represented, data collection method, year(s) of data collection, sex and age range, diagnostic criteria or measurement method, and sample size, as relevant.	Supplementary Information: 3.0 Supplementary data
6	Identify and describe any categories of input data that have potentially important biases (e.g., based on characteristics listed in item 5).	Main text: Discussion (Limitations)
For data	inputs that contribute to the analysis but were not synthesized as part of the study:	
7	Describe and give sources for any other data inputs.	Main text: Methods (Geospatial covariates), Supplementary Information: 4.0 Supplementary covariates
For all de	ata inputs:	
8	Provide all data inputs in a file format from which data can be efficiently extracted (e.g., a spreadsheet rather than a PDF), including all relevant meta- data listed in item 5. For any data inputs that cannot be shared because of ethical or legal reasons, such as third-party ownership, provide a contact name or the name of the institution that retains the right to the data.	Available at (GHDx link will be added upon review) Supplementary Information: 3.0 Supplementary data
9	Provide a conceptual overview of the data analysis method. A diagram may be helpful.	Main text: Methods (Age and diagnostic adjustment), Supplementary Information: 5.1 Age and diagnostic crosswalks; Supplementary Information: Table 6: data used in estimation of age and diagnostic crosswalk
10	Provide a detailed description of all steps of the analysis, including mathematical formulae. This description should cover, as relevant, data cleaning, data pre-processing, data adjustments and weighting of data sources, and mathematical or statistical model(s).	Main text: Methods (Geospatial analysis), Supplementary Information: 5.3 Geostatistical model
11	Describe how candidate models were evaluated and how the final model(s) were selected.	Main text: Methods (Geospatial analysis), Supplementary Information: 5.4 Model validation
12	Provide the results of an evaluation of model performance, if done, as well as the results of any relevant sensitivity analysis.	Main text: Methods (Geospatial analysis), Supplementary Information: 5.4 Model validation
13	Describe methods for calculating uncertainty of the estimates. State which sources of uncertainty were, and were not, accounted for in the uncertainty analysis.	Main text: Methods (Geospatial analysis), Supplementary Information: 5.3.4. Model description
14	State how analytic or statistical source code used to generate estimates can be accessed.	Available at (GHDx link will be added upon review)
Results a	nd Discussion	
15	Provide published estimates in a file format from which data can be efficiently extracted.	Raster files for spatial data and CSVs of estimates available at (GHDx link will be added upon review)
16	Report a quantitative measure of the uncertainty of the estimates (e.g., uncertainty intervals)	Supplementary Information: Supplementary Figure 11 and Table 9

17	Interpret results in light of existing evidence. If updating a previous set of estimates, describe the reasons for changes in estimates.	Main text: Discussion (Strengths)
18	Discuss limitations of the estimates. Include a discussion of any modelling assumptions or data limitations that affect interpretation of the estimates.	Main text: Discussion (Limitations)

70 2.0 Supplementary discussion

71 This document outlines the major data processing, modelling, and validation steps for the onchocerciasis prevalence

analysis described in the main text (Supplementary Figure 1). We present a detailed description of model inputs,

including data coverage, covariate sources, and geo-referencing. The geospatial model is described along with model validation metrics.



Supplementary Figure 1. Flowchart of major steps in data processing and modelling of onchocerciasis prevalence.

79 **3.0 Supplementary data**

80 In the following section, we present a detailed summary of the data inputs used to estimate the prevalence of

onchocerciasis in Africa and Yemen. Broadly, we aimed to include all published sources of onchocerciasis infection
 prevalence, as well as routine programme monitoring data collected to monitor progress toward onchocerciasis

elimination. Data inputs were retained for analysis if they could be accurately geo-referenced.

83 84

85 **3.1 Geographical restrictions**

- 86 Supplementary Table 2 lists all countries included in our MBG (model-based geostatistical) modelling region.
- 87 Inclusion was partially based on ESPEN (Expanded Special Project for Elimination of Neglected Tropical Diseases)

- 88 onchocerciasis endemicity classifications,(1) with extensions to some neighbouring countries outside the WHO
- (World Health Organization) AFRO (Regional Office for Africa) region which have evidence of onchocerciasis
 endemicity or uncertain endemicity status (Sudan, Somalia, and Yemen). Portions of Mauritius, Namibia, and
- 90 endemicity or uncertain endemicity status (Sudan, Somalia, and Yemen). Portions of Mauritius, Namibia, and 91 Zambia are considered by ESPEN as candidates for future elimination mapping but were not included in our
- 91 Zambia are considered by ESPEN as candidates for future elimination mapping but were not included in our 92 geospatial model. Conversely, The Gambia is considered non-endemic but we included it in our model for
- 92 geospatial model. Conversely, The Gambia is considered non-endemic but we included it in our model for 93 geographical continuity. Countries in the Americas with historical or residual onchocerciasis burdens were not
- 94 modelled due to their highly localised endemicity.

95 Supplementary Table 2. Geographical restrictions.

96 The geographical definition of the modelling region is indicated, with country-level onchocerciasis endemicity

97 status per ESPEN, and which countries were included in our geospatial analysis from 2000 to 2018. Endemicity

98 classifications were derived from ESPEN data at the level of intervention units (IU) (retrieved 14 February 2020).

99 Countries outside the WHO AFRO region are not covered by the scope of ESPEN; countries in the WHO EMRO

ESPEN endemicity

100 (Eastern Mediterranean Regional Office) region are listed here if they are in Africa or have evidence of 101 onchocerciasis endemicity or uncertain status. We considered countries endemic if they had at least one intervent

onchocerciasis endemicity or uncertain status. We considered countries endemic if they had at least one intervention
 unit that was flagged as endemic by ESPEN. Number of observations: The number of data rows in the final dataset
 for a given country.

Included in 2000–2018 model?

103 104

Location

Observations AFRO(1) Algeria Non-endemic no Angola Endemic 876 ves Benin Endemic 300 yes Botswana Non-endemic no Burkina Faso Endemic 72 yes Burundi Endemic 263 yes Cabo Verde Non-endemic no Cameroon Endemic 1216 yes Central African Republic Endemic 1079 yes Chad Endemic 753 yes Comoros Non-endemic no Republic of the Congo Endemic yes 490 Côte d'Ivoire Endemic 95 yes Democratic Republic of the Endemic 4574 yes Congo Equatorial Guinea Endemic 341 yes Eritrea Non-endemic no Eswatini Non-endemic no

Number of

Location	ESPEN endemicity	Included in 2000–2018 model?	Number of Observations
Ethiopia	Endemic	yes	1062
Gabon	Endemic	yes	157
The Gambia	Non-endemic	yes	0
Ghana	Endemic	yes	78
Guinea	Endemic	yes	193
Guinea-Bissau	Endemic	yes	0
Kenya	Consider oncho elimination mapping	yes	94
Lesotho	Non-endemic	no	-
Liberia	Endemic	yes	114
Madagascar	Non-endemic	no	-
Malawi	Endemic	yes	333
Mali	Endemic	yes	160
Mauritania	Non-endemic	no	-
Mauritius	Mapping gap (one district)	no	-
Mozambique	Endemic	yes	291
Namibia	Consider oncho elimination mapping (some districts)	no	-
Niger	Endemic	yes	0
Nigeria	Endemic	yes	3445
Rwanda	Consider oncho elimination mapping	yes	90
São Tomé and Príncipe	Non-endemic	yes	0
Senegal	Endemic	yes	67
Seychelles	Non-endemic	no	-
Sierra Leone	Endemic	yes	223
South Africa	Non-endemic	no	-
South Sudan	Endemic	yes	466
Togo	Endemic	yes	160
Uganda	Endemic	yes	620
Tanzania	Endemic	yes	478

Location	ESPEN endemicity	Included in 2000–2018 model?	Number of Observations
Zambia	Consider oncho elimination mapping (some districts)	no	-
Zimbabwe	Non-endemic	no	-
<i>EMRO</i> (2)			
Djibouti	-	no	-
Egypt	-	no	-
Libya	-	no	-
Morocco	-	no	-
Somalia	-	yes	0
Sudan	(Endemic)	yes	26
Tunisia	-	no	-
Yemen	(Endemic)	yes	0

106

107 **3.2** Systematic review

Articles related to onchocerciasis were found by searching PubMed, Scopus, and Web of Science using the following keywords: "Oncho", "river blindness", "O. Volvulus", "robles disease", "blinding filariasis", "coast

109 following keywords: "Oncho", "fiver blindness", "O. volvulus", "robles disease", "blinding filariasis", "coas 110 erysipelas", and "sowda". A systematic review of these reports, all published before July 7, 2017, was then

111 conducted. A second round of formal reviews was undertaken on June 6, 2019, to cover new articles published after

112 July 7, 2017, and followed the same process as the first round.

113

114 3.2.1 Systematic review data processing

115 The systematic review process is illustrated in Supplementary Figure 2. Throughout the systematic review we 116 excluded publications that met the following criteria: no measurement of onchocerciasis prevalence, data collected 117 before 1985, case-control studies, qualitative research publications, duplicative data from cohort studies, hospital-118 based studies, and publications that did not report the location of data collection. The search identified 4048 119 publications, which were reduced to 579 after screening titles and abstracts. A full-text review was completed for the 120 remaining publications. The full-text review yielded 259 publications which met the inclusion criteria and were 121 extracted. The literature review was updated on June 6, 2019, by searching PubMed with the same search string used 122 in 2017 for articles published after July 7, 2017. The search returned 304 results, which were narrowed to 115 123 articles after screening titles and abstracts. A full-text review resulted in 22 articles that were eligible for extraction. 124 Additional publications were identified outside of the literature review and screened for inclusion by the same 125 criteria, up through April 14, 2020. The final dataset drew from 290 articles. Overall, 16 096 datapoints were 126 extracted and geo-located from 290 publications. Data extracted from each source included survey year, age-range

of individuals tested, diagnostic information, sample size, number of individuals tested positive, and sampling
 strategy details.



31 Supplementary Figure 2. Onchocerciasis article review and data extraction flowchart.

132 Each step of the extraction process is outlined, from article identification and screening to extraction, including the

133 number of articles or records that were processed or removed in each step before reaching the final dataset.

134 Additional articles outside of the literature review were identified and screened for inclusion ("ongoing process") on

- an ongoing basis up until April 14, 2020.
- 136

137 **3.3 Geo-positioning**

138 Geographical information associated with onchocerciasis prevalence data were verified and geo-located to ensure 139 accuracy. Data associated with locations smaller than 5×5 km were treated as points and geo-located as

139 accuracy. Data associated with locations smaller than 5×5 km were treated as points and geo-located as 140 latitude/longitude coordinates. Coordinates provided by a data source were mapped to ensure that the coordinates

were located in the correct administrative units. If coordinates were not reported, points were geolocated and vetted

by comparing results from Google Maps, Fuzzy Gazetteer, Geonames, and Open Street Map. Locations larger than 5

142 by comparing results from Google Waps, Fuzzy Gazetteer, Geonanes, and Open Street Wap. Eccations farger than 143 × 5 km were treated as polygon data and geo-located to the most granular administrative boundary possible (most

144 commonly district level). Custom shapefiles were created to geolocate areas that did not align with administrative

- boundaries. In the event that a literature source only included a map of locations sampled without any other
- 146 information, ArcGIS software was utilised to overlay the map onto existing administrative boundaries, and location
- 147 coordinates or custom polygons were manually created and recorded. Prevalence data that were reported for
- administrative areas were matched to their appropriate polygon by searching our administrative shapefile database.
- 149 If place names were unidentifiable across multiple shapefile libraries or geo-referencing sources, they were excluded
- 150 from the analysis.
- 151

152 **3.4 Data processing**

153 Data were excluded from the analysis for the following reasons: did not report survey year (N = 8236); did not

report post-1988 data (N = 2648); did not report skin snip microscopy or nodule prevalence (N = 12 121); did not

- report sample size or prevalence (N = 30); or did not report data for countries in the modelling region or could not
- be accurately georeferenced (N = 580). Duplicate records were identified and excluded (N = 1592); these generally
- 157 reflected different data sources (eg, literature extraction and ESPEN) which contained the same survey, as judged by
- 158 identical year, location, sample size, cases, and reference when available, although there was some deduplication
- within individual data sources. Prior to polygon resampling (see section 5.2), the dataset consisted of 18 116
- 160 georeferenced prevalence observations, consisting of 17 896 point-referenced observations and 220 areal
- 161 observations. A total of 14 314 observations represented prevalence based on nodule examinations, and 3802
- 162 observations represented microfiladermia prevalence as measured by skin snip assays. After resampling, the full
- 163 modelling dataset consisted of 20 124 georeferenced datapoints.
- 164
- 165 Supplementary Table 3 provides citations for data sources used in our onchocerciasis MBG model. The
- 166 geographical coverage of the final dataset is summarised in Supplementary Figure 3.

167 Supplementary Table 3. Citations for data inputs.

168 The NID is a unique identifier cataloguing all data inputs in the Global Health Data Exchange

169 170 171 (http://ghdx.healthdata.org). Note: Records are listed here in alphabetical order by geography, but some sources

provided data for multiple countries; such sources are listed here only once.

NID	Geographies	Citation
332798	Angola	Carme B, Ntsoumou-Madzou V, Samba Y, Yebakima A. Prevalence of depigmentation of the shins: a simple and cheap way to screen for severe endemic onchocerciasis in Africa. Bull World Health Organ. 1993; 71(6): 755–8.
334477	Benin	Gallin M, Adams A, Kruppa TF, Gbaguidi EA, Massougbodji A, Sadeler BC, Brattig N, Erttmann KD. Epidemiological studies of onchocerciasis in southern Benin. Trop Med Parasitol. 1993; 44(2): 69–74.
125405	Burkina Faso	Kabore JK, Cabore JW, Melaku Z, Druet-Cabanac M, Preux PM. Epilepsy in a focus of onchocerciasis in Burkina Faso. Lancet. 1996; 347(9004): 836.
334475	Burkina Faso	De Sole G, Remme J. Importance of migrants infected with Onchocerca volvulus in west African river valleys protected by 14 to 15 years of Simulium control. Trop Med Parasitol. 1991; 42(2): 75–8.
332903	Burkina Faso, Côte d'Ivoire	Toè L, Adjami AG, Boatin BA, Back C, Alley ES, Dembélé N, Brika PG, Pearlman E, Unnasch TR. Topical application of diethylcarbamazine to detect onchocerciasis recrudescence in west Africa. Trans R Soc Trop Med Hyg. 2000; 94(5): 519–25.
334481	Burundi	Newell E d. Comparison of the use of skin scarification and skin biopsies to determine the prevalence and intensity of Onchocerca volvulus infection. Ann Trop Med Parasitol. 1997; 91(6): 633.
338571	Burundi	Newell ED, Hicuburundi B, Ndimuruvugo N. [Endemicity and clinical manifestations of onchocerciasis in the province of Bururi, Burundi]. Trop Med Int Health. 1997; 2(3): 218–26.
338573	Burundi	Newell ED, Ndimuruvugo N, Nimpa D. [Endemicity and clinical manifestations of onchocerciasis in the provinces of Cibitoke and Bubanza (Burundi)]. Bull Soc Pathol Exot. 1997; 90(5): 353–7.
136492	Cameroon	Cho-Ngwa F, Amambua AN, Ambele MA, Titanji VPK. Evidence for the exacerbation of lymphedema of geochemical origin, podoconiosis, by onchocerciasis. J Infect Public Health. 2009; 2(4): 198-203.
159316	Cameroon	Matthews GA, Dobson HM, Nkot PB, Wiles TL, Birchmore M. Preliminary examination of integrated vector management in a tropical rainforest area of Cameroon. Trans R Soc Trop Med Hyg. 2009; 103(11): 1098-104.
324729	Cameroon	Kamga G-R, Dissak-Delon FN, Nana-Djeunga HC, Biholong BD, Mbigha- Ghogomu S, Souopgui J, Zoure HGM, Boussinesq M, Kamgno J, Robert A. Still mesoendemic onchocerciasis in two Cameroonian community-directed treatment with ivermectin projects despite more than 15 years of mass treatment. Parasites Vectors. 2016; 9(1): 581.
327960	Cameroon	Wanji S, Kengne-Ouafo JA, Esum ME, Chounna PWN, Tendongfor N, Adzemye BF, Eyong JEE, Jato I, Datchoua-Poutcheu FR, Kah E, Enyong P, Taylor DW. Situation analysis of parasitological and entomological indices of onchocerciasis transmission in three drainage basins of the rain forest of South West Cameroon after a decade of ivermectin treatment. Parasites Vectors. 2015; 8: 202.
327968	Cameroon	Katabarwa MN, Eyamba A, Chouaibou M, Enyong P, Kuété T, Yaya S, Yougouda A, Baldiagaï J, Madi K, Andze GO, Richards F. Does onchocerciasis transmission take place in hypoendemic areas? a study from the North Region of Cameroon. Trop Med Int Health. 2010; 15(5): 645-52.

NID	Geographies	Citation
327992	Cameroon	Katabarwa MN, Eyamba A, Nwane P, Enyong P, Kamgno J, Kueté T, Yaya S, Aboutou R, Mukenge L, Kafando C, Siaka C, Mkpouwoueiko S, Ngangue D, Biholong BD, Andze GO. Fifteen years of annual mass treatment of onchocerciasis with ivermectin have not interrupted transmission in the west region of cameroon. J Parasitol Res. 2013; 2013: 420928.
328065	Cameroon	Katabarwa MN, Eyamba A, Nwane P, Enyong P, Yaya S, BaldiagaÃ ⁻ J, Madi TK, Yougouda A, Andze GO, Richards FO. Seventeen years of annual distribution of ivermectin has not interrupted onchocerciasis transmission in North Region, Cameroon. Am J Trop Med Hyg. 2011; 85(6): 1041-9.
328095	Cameroon	Kamga HLF, Shey DN, Assob JCN, Njunda AL, Nde Fon P, Njem PK. Prevalence of onchocerciasis in the Fundong Health District, Cameroon after 6 years of continuous community-directed treatment with ivermectin. Pan Afr Med J. 2011; 10: 34.
332745	Cameroon	Pion SDS, Clarke P, Filipe J a. N, Kamgno J, Gardon J, Basáñez M-G, Boussinesq M. Co-infection with Onchocerca volvulus and Loa loa microfilariae in central Cameroon: are these two species interacting?. Parasitology. 2006; 132(Pt 6): 843–54.
332783	Cameroon	Boussinesq M, Chippaux JP, Ernould JC, Quillevere D, Prod'hon J. Effect of repeated treatments with ivermectin on the incidence of onchocerciasis in northern Cameroon. Am J Trop Med Hyg. 1995; 53(1): 63–7.
332786	Cameroon	Kollo B, Mather FJ, Cline BL. Evaluation of alternate methods of rapid assessment of endemicity of Onchocerca volvulus in communities in southern Cameroon. Am J Trop Med Hyg. 1995; 53(3): 243–7.
332788	Cameroon	Ngoumou P, Walsh JF, Mace JM. A rapid mapping technique for the prevalence and distribution of onchocerciasis: a Cameroon case study. Ann Trop Med Parasitol. 1994; 88(5): 463–74.
332790	Cameroon	Mendoza Aldana J, Piechulek H, Maguire J. Forest onchocerciasis in Cameroon: its distribution and implications for selection of communities for control programmes. Ann Trop Med Parasitol. 1997; 91(1): 79–86.
332840	Cameroon	Ayong LS, Tume CB, Wembe FE, Simo G, Asonganyi T, Lando G, Ngu JL. Development and evaluation of an antigen detection dipstick assay for the diagnosis of human onchocerciasis. Trop Med Int Health. 2005; 10(3): 228-33.
332848	Cameroon	Wanji S, Tendongfor N, Esum M, Ndindeng S, Enyong P. Epidemiology of concomitant infections due to Loa loa, Mansonella perstans, and Onchocerca volvulus in rain forest villages of Cameroon. Med Microbiol Immunol. 2003; 192(1): 15-21.
332893	Cameroon	Kamgno J, Boussinesq M. [Hyperendemic loaiasis in the Tikar plain, shrub savanna region of Cameroon]. Bull Soc Pathol Exot. 2001; 94(4): 342–6.
332897	Cameroon	Esum M, Wanji S, Tendongfor N, Enyong P. Co-endemicity of loiasis and onchocerciasis in the South West Province of Cameroon: implications for mass treatment with ivermectin. Trans R Soc Trop Med Hyg. 2001; 95(6): 673–6.
338584	Cameroon	Kamgno J, Bouchité B, Baldet T, Folefack G, Godin C, Boussinesq M. [Study of the distribution of human filariasis in West Province of Cameroon]. Bull Soc Pathol Exot. 1997; 90(5): 327–30.
332820	Cameroon, Central African Republic, Gabon	Ozoh G, Boussinesq M, Bissek A-CZ-K, Kobangue L, Kombila M, Mbina J-RM, Enyong P, Noma M, Sékétéli A, Fobi G. Evaluation of the diethylcarbamazine patch to evaluate onchocerciasis endemicity in Central Africa. Trop Med Int Health. 2007; 12(1): 123-9.

NID	Geographies	Citation
327872	Cameroon, Nigeria, Sudan, Uganda	Ozoh GA, Murdoch ME, Bissek A-C, Hagan M, Ogbuagu K, Shamad M, Braide EI, Boussinesq M, Noma MM, Murdoch IE, Sékétéli A, Amazigo UV. The African Programme for Onchocerciasis Control: impact on onchocercal skin disease. Trop Med Int Health. 2011; 16(7): 875–83.
332855	Central African Republic	Kennedy MH, Bertocchi I, Hopkins AD, Meredith SE. The effect of 5 years of annual treatment with ivermectin (Mectizan) on the prevalence and morbidity of onchocerciasis in the village of Gami in the Central African Republic. Ann Trop Med Parasitol. 2002; 96(3): 297-307.
332796	Congo	Brito M, Paulo R, Van-Dunem P, Martins A, Unnasch TR, Novak RJ, Jacob B, Stanton MC, Molyneux DH, Kelly-Hope LA. Rapid Integrated Clinical Survey to Determine Prevalence and Co-distribution Patterns of Lymphatic Filariasis and Onchocerciasis in a Loa Loa Co-endemic Area: The Angolan Experience. Parasite Epidemiol Control. 2017; 2(3): 71-84.
338569	Congo	Talani P, Missamou F, Kankou JM, Niabe B, Obengui null, Moyen G. [Rapid epidemiological mapping of onchocerciasis in Congo Brazzaville]. Dakar Med. 2005; 50(3): 164–7.
327962	Côte d'Ivoire	Lloyd MM, Gilbert R, Taha NT, Weil GJ, Meite A, Kouakou IMM, Fischer PU. Conventional parasitology and DNA-based diagnostic methods for onchocerciasis elimination programmes. Acta Trop. 2015; 146: 114-8.
338588	Côte d'Ivoire	Diarrassouba S, Traore S, Riviere F. [Endemic onchocerciasis in forested zones of Ivory Coast: prevalence rate and microfilarial densities]. Med Trop (Mars). 1996; 56(1): 59–62.
332853	Democratic Republic of Congo	Kayembe DL, Kasonga DL, Kayembe PK, Mwanza J-CK, Boussinesq M. Profile of eye lesions and vision loss: a cross-sectional study in Lusambo, a forest-savanna area hyperendemic for onchocerciasis in the Democratic Republic of Congo. Trop Med Int Health. 2003; 8(1): 83-9.
327885	Ethiopia	Feleke SM, Tadesse G, Mekete K, Tekle AH, Kebede A. Epidemiological Mapping of Human Onchocerciasis in Transmission Suspected Districts of Bale, Borena, and West Arsi Zones of Eastern Ethiopia. Interdiscip Perspect Infect Dis. 2016; 2016: 6937509.
327957	Ethiopia	Dana D, Debalke S, Mekonnen Z, Kassahun W, Suleman S, Getahun K, Yewhalaw D. A community-based cross-sectional study of the epidemiology of onchocerciasis in unmapped villages for community directed treatment with ivermectin in Jimma Zone, southwestern Ethiopia. BMC Public Health. 2015; 15: 595.
328057	Ethiopia	Dori GU, Belay T, Belete H, Panicker KN, Hailu A. Parasitological and clinico- epidemiological features of onchocerciasis in West Wellega, Ethiopia. J Parasit Dis. 2012; 36(1): 10-8.
332186	Ethiopia	Mengistu G, Balcha F, Britton S. Co-infection of Onchocerca volvulus and intestinal helminths in indigenous and migrant farmers in southwest Ethiopia. Ethiop Med J. 2002; 40(1): 19–27.
332189	Ethiopia	Jira C. Prevalence of onchocerciasis in Blue Nile valley of western Ethiopia. Indian J Public Health. 1993; 37(4): 135–7.
332193	Ethiopia	Taye A, Gebre-Michael T, Taticheff S. Onchocerciasis in Gilgel Ghibe River Valley southwest Ethiopia. East Afr Med J. 2000; 77(2): 116–20.
332674	Ethiopia	Adugna N, Woldegiorgis M, Tilahun D, Kebede A, Hadis M. The Epidemiology of Onchocerciasis in the Resettled and Indigenous Population in Pawe, Western Ethiopia. Ethiop Med J. 1999; 37(1): 41-50.

NID	Geographies	Citation
332676	Ethiopia	Legesse M, Balcha F, Erko B. Status of onchocerciasis in Teppi area, Southwestern Ethiopia, after four years of annual community-directed treatment with ivermectin. Ethiop J Health Dev. 2010; 24(1): 51-56.
332678	Ethiopia	Samuel A, Belay T, Yehalaw D, Taha M, Zemene E, Zeynudin A. Impact of Six Years Community Directed Treatment with Ivermectin in the Control of Onchocerciasis, Western Ethiopia. PLoS One. 2016; 11(3): e0141029.
332851	Ethiopia	Hailu A, Balcha F, Birrie H, Berhe N, Aga A, Mengistu G, Bezuneh A, Ali A, Gebre-Michael T, Gemetchu T. Prevalence of onchocercal skin disease and infection among workers of coffee plantation farms in Teppi, southwestern Ethiopia. Ethiop Med J. 2002; 40(3): 259-69.
332825	Gabon	Fobi G, Mourou Mbina JR, Ozoh G, Kombila M, Agaya C, Olinga Olinga JM, Boussinesq M, Enyong P, Noma M, Sékétéli A. [Onchocerciasis in the area of Lastourville, Gabon. Clinical and entomological aspects]. Bull Soc Pathol Exot. 2006; 99(4): 269-71.
327974	Ghana	Garms R, Badu K, Owusu-Dabo E, Baffour-Awuah S, Adjei O, Debrah AY, Nagel M, Biritwum NK, Gankpala L, Post RJ, Kruppa TF. Assessments of the transmission of Onchocerca volvulus by Simulium sanctipauli in the Upper Denkyira District, Ghana, and the intermittent disappearance of the vector. Parasitol Res. 2015; 114(3): 1129-37.
327912	Ghana, Sri Lanka	Frempong KK, Walker M, Cheke RA, Tetevi EJ, Gyan ET, Owusu EO, Wilson MD, Boakye DA, Taylor MJ, Biritwum N-K, Osei-Atweneboana M, Basáñez M-G. Does Increasing Treatment Frequency Address Suboptimal Responses to Ivermectin for the Control and Elimination of River Blindness?. Clin Infect Dis. 2016; 62(11): 1338–47.
332662	Malawi, Tanzania	Courtright P, Johnston K, Chitsulo L. A new focus of onchocerciasis in Mwanza District, Malawi. Trans R Soc Trop Med Hyg. 1995; 89(1): 34–6.
144330	Mali, Senegal	Diawara L, Traoré MO, Badji A, Bissan Y, Doumbia K, Goita SF, Konaté L, Mounkoro K, Sarr MD, Seck AF, Toé L, Tourée S, Remme JH. Feasibility of onchocerciasis elimination with ivermectin treatment in endemic foci in Africa: first evidence from studies in Mali and Senegal. PLoS Negl Trop Dis. 2009; 3(7): e497.
191825	Mali, Senegal	Traore MO, Sarr MD, Badji A, Bissan Y, Diawara L, Doumbia K, Goita SF, Konate L, Mounkoro K, Seck AF, Toe L, Toure S, Remme JH. Proof-of-principle of onchocerciasis elimination with ivermectin treatment in endemic foci in Africa: final results of a study in Mali and Senegal. PLoS Negl Trop Dis. 2012; 6(9): e1825.
136520	Nigeria	Anosike JC, Onwuliri CO. Studies on filariasis in Bauchi State, Nigeria. II. The prevalence of human filariasis in Darazo Local Government area. Appl Parasitol. 1994; 35(4): 242-50.
147655	Nigeria	Anosike JC, Onwuliri COE, Onwuliri VA. Human filariasis in Dass local government area of Bauchi state, Nigeria. Trop Ecol. 2003; 44(2): 217-27.
147869	Nigeria	Uttah E, Ibeh DC. Multiple filarial species microfiladermia: a comparative study of areas with endemic and sporadic onchocerciasis. J Vector Borne Dis. 2011; 48(4): 197-204.
324133	Nigeria	Evans DS, Alphonsus K, Umaru J, Eigege A, Miri E, Mafuyai H, Gonzales-Peralta C, Adamani W, Pede E, Umbugadu C, Saka Y, Okoeguale B, Richards FO. Status of Onchocerciasis transmission after more than a decade of mass drug administration for onchocerciasis and lymphatic filariasis elimination in central Nigeria: challenges in coordinating the stop MDA decision. PLoS Negl Trop Dis. 2014; 8(9): 1-10.

NID	Geographies	Citation
327876	Nigeria	Eyo J, Onyishi G, Ugokwe C. Rapid epidemiological assessment of onchocerciasis in a tropical semi-urban community, enugu state, Nigeria. Iran J Parasitol. 2013; 8(1): 145–51.
327921	Nigeria	Katabarwa M, Eyamba A, Habomugisha P, Lakwo T, Ekobo S, Kamgno J, Kuete T, Ndyomugyenyi R, Onapa A, Salifou M, Ntep M, Richards FO. After a decade of annual dose mass ivermectin treatment in Cameroon and Uganda, onchocerciasis transmission continues. Trop Med Int Health. 2008; 13(9): 1196-1203.
327941	Nigeria	Okoye IC, Onwuliri CO. Epidemiology and psycho-social aspects of onchocercal skin diseases in northeastern Nigeria. Filaria J. 2007; 6: 15.
327954	Nigeria	Okoye IC, Onwuliri CO. Epidemiology and psycho-social aspects of onchocercal skin diseases in northeastern Nigeria. Filaria J. 2007; 6: 15.
327966	Nigeria	Ojurongbe O, Akindele AA, Adeleke MA, Oyedeji MO, Adedokun SA, Ojo JF, Akinleye CA, Bolaji OS, Adefioye OA, Adeyeba OA. Co-endemicity of loiasis and onchocerciasis in rain forest communities in southwestern Nigeria. PLoS Negl Trop Dis. 2015; 9(3): e0003633.
327972	Nigeria	Opara KN, Fagbemi BO. Population dynamics of onchocerca volvulus microfilariae in human host after six years of drug control. J Vector Borne Dis. 2008; 45(1): 29-37.
327982	Nigeria	Akinbo FO, Okaka CE. Hyperendemicity of onchocerciasis in Ovia Northeast Local Government Area, Edo State, Nigeria. East Afr J Public Health. 2010; 7(1): 84-6.
328059	Nigeria	Osue HO, Inabo HI, Yakubu SE, Audu PA, Galadima M, Odama LE, Musa D, Ado SA, Mamman M. Impact of Eighteen-Year Varied Compliance to Onchocerciasis Treatment with Ivermectin in Sentinel Savannah Agrarian Communities in Kaduna State of Nigeria. ISRN Parasitol. 2013; 2013: 960168.
328063	Nigeria	Tekle AH, Elhassan E, Isiyaku S, Amazigo UV, Bush S, Noma M, Cousens S, Abiose A, Remme JH. Impact of Long-term Treatment of Onchocerciasis with Ivermectin in Kaduna State, Nigeria: First Evidence of the Potential for Elimination in the Operational Area of the African Programme for Onchocerciasis Control. Parasites Vectors. 2012; 5(1).
328083	Nigeria	Uttah E. Onchocerciasis in the upper imo river basin, Nigeria: prevalence and comparative study of waist and shoulder snips from mesoendemic communities. Iran J Parasitol. 2010; 5(2): 33-41.
328089	Nigeria	Sam-Wobo SO, Adeleke MA, Jayeola OA, Adeyi AO, Oluwole AS, Ikenga M, Lawniye F, Gazama J, Kagni A, Kosoko TO, Agbeyangi O, Bankole S, Toé L, Mafiana CF, Yameogo L. Epidemiological evaluation of onchocerciasis along Ogun River System, southwest Nigeria. J Vector Borne Dis. 2012; 49(2): 101-4.
332571	Nigeria	Akogun OB, Akoh JI, Hellandendu H. Non-ocular clinical onchocerciasis in relation to skin microfilaria in the Taraba River Valley, Nigeria. J Hyg Epidemiol Microbiol Immunol. 1992; 36(4): 368–83.
332576	Nigeria	Anosike JC, Onwuliri CO. Studies on filariasis in Bauchi State, Nigeria. 1. Endemicity of human onchocerciasis in Ningi Local Government Area. Ann Trop Med Parasitol. 1995; 89(1): 31–8.
332594	Nigeria	Nwaorgu OC, Ohaegbula A, Onweluzo IE, Alo ET, Nweke LN, Agu ML, Emeh E. Results of a large scale onchocercosis survey in Enugu State, Nigeria. J Helminthol. 1994; 68(2): 155–9.
332703	Nigeria	Emukah E, Nimzing J, Ezeabikwa M, Obiezu J, Okpala N, Miri E, Katabarwa M, Hopkins D, Richards F. Impact assessment of repeated annual ivermectin on ocular

NID	Geographies	Citation
		and clinical onchocerciasis 14 years of annual mass drug administration in eight sentinel villages, southeast Nigeria 2008. Am J Trop Med Hyg. 2010; 83(Suppl 5): 22.
332707	Nigeria	Olofintoye LK, Adewole SO, Erinle FB, Ajayi AI. Prevalence of Onchoceriasis Syndrome in Ise-Orun Local Government Area of Ekiti State, Nigeria. Pak J Sci Ind Res. 2005; 48(6): 412-6.
332712	Nigeria	Oyibo WA, Fagbenro-Beyioku AF. Reduced prevalence of onchocerciasis following mass treatment with ivermectin. East Afr Med J. 1997; 74(5): 326–30.
332714	Nigeria	Rebecca SN, Akinboye DO, Abdulazeez AA. Onchocerciasis and Plasmodiasis: Concurrent Infection in Garaha-Dutse Community, Adamawa State Nigeria. Biomed Res. 2008; 19(2): 107-11.
332717	Nigeria	Abanobi OC, Anosike JC. Control of onchocerciasis in Nzerem-Ikpem, Nigeria: baseline prevalence and mass distribution of ivermectin. Public Health. 2000; 114(5): 402–6.
332719	Nigeria	Akinboye DO, Okwong E, Ajiteru N, Fawole O, Agbolade O, Ayinde OO, Amosu AM, Atulomah NOS, Oduola O, Owodunni BM, Rebecca SN, Falade M. Onchocerciasis Among Inhabitants of Ibarapa Local Government Community of Oyo State, Nigeria. Biomed Res. 2010; 21(2): 174-8.
332723	Nigeria	Akogun OB, Musa-Hong H, Hellandendu H. Onchocerciasis in Taraba State, Nigeria: intensity, rate of infection and associated symptoms in 14 communities of Bali district. Appl Parasitol. 1994; 35(2): 125–32.
332725	Nigeria	Akogun OB, Akoh JI, Okolo A. Comparison of two sample survey methods for hyperendemic onchocerciasis and a new focus in Dakka, Nigeria. Rev Biol Trop. 1997; 45(2): 871–6.
332728	Nigeria	Akogun OB. Onchocerciasis in Taraba State, Nigeria: clinical-epidemiological study of at-risk males in Bakundi District. Zentralbl Bakteriol. 1999; 289(3): 371–9.
332730	Nigeria	Nmorsi OPG, Oladokun I a. A, Egwunyenga OA, Oseha E. Eye lesions and onchocerciasis in a rural farm settlement in Delta state, Nigeria. Southeast Asian J Trop Med Public Health. 2002; 33(1): 28–32.
332732	Nigeria	Opara KN, Fagbemi BO, Atting IA, Oyene UE, Okenu DMN. Status of forest onchocerciasis in the Lower Cross River Basin, Nigeria: change in clinical and parasitological indices after 6 years of ivermectin intervention. Public Health. 2007; 121(3): 202–7.
332734	Nigeria	Umeh RE, Chijioke CP, Okonkwo PO. Eye disease in an onchocerciasis-endemic area of the forest-savanna mosaic region of Nigeria. Bull World Health Organ. 1996; 74(1): 95–100.
332736	Nigeria	Umeh RE. The causes and profile of visual loss in an onchocerciasis-endemic forest-savanna zone in Nigeria. Ophthalmic Epidemiol. 1999; 6(4): 303–15.
332899	Nigeria	Anosike JC, Celestine NA, Onwuliri OE, Onwuliri VA. The prevalence, intensity and clinical manifestations of Onchocerca volvulus infection in Toro local government area of Bauchi State, Nigeria. Int J Hyg Environ Health. 2001; 203(5– 6): 459–64.
332905	Nigeria	Nwosu SN. Low vision in persons aged 50 and above in the onchocercal endemic communities of Anambra State, Nigeria. West Afr J Med. 2000; 19(3): 216–9.

NID	Geographies	Citation
332907	Nigeria	Abanobi OC. Cross-validation of the rapid epidemiological mapping of onchocerciasis endemicity in Anambra state, Nigeria. Ann Trop Med Parasitol. 1999; 93(7): 721–6.
327879	Senegal	Wilson NO, Badara Ly A, Cama VA, Cantey PT, Cohn D, Diawara L, Direny A, Fall M, Feeser KR, Fox LM, Kabore A, Seck AF, Sy N, Ndiaye D, Dubray C. Evaluation of Lymphatic Filariasis and Onchocerciasis in Three Senegalese Districts Treated for Onchocerciasis with Ivermectin. PLoS Negl Trop Dis. 2016; 10(12): e0005198.
136415	Sierra Leone	Gbakima AA, Sahr F. Filariasis in the Kaiyamba Chiefdom, Moyamba District Sierra Leone: an epidemiological and clinical study. Public Health. 1996; 110(3): 169-74.
155864	Sierra Leone	Gbakima AA. Inland valley swamp rice development: malaria, schistosomiasis, onchocerciasis in south central Sierra Leone. Public Health. 1994; 108(2): 149-57.
334430	Sierra Leone	Gbakima AA, Barbe RF. Onchocerca Volvulus infection in Sierra Leone: relation between prevalence, intensity of infection, and ocular problems in a "forest" region. Acta Leiden. 1992; 60(2): 47–59.
334432	Sierra Leone	Gbakima AA. Integrated control of Onchocerca volvulus infection in a hyperendemic zone in Sierra Leone. East Afr Med J. 1996; 73(3): 159–63.
327874	Sudan	Higazi TB, Zarroug IMA, Mohamed HA, Elmubark WA, Deran TCM, Aziz N, Katabarwa M, Hassan HK, Unnasch TR, Mackenzie CD, Richards F, Hashim K. Interruption of Onchocerca volvulus transmission in the Abu Hamed focus, Sudan. Am J Trop Med Hyg. 2013; 89(1): 51–7.
327986	Tanzania	Mweya CN, Kalinga AK, Kabula B, Malley KD, Ruhiso MH, Maegga BTA. Onchocerciasis situation in the Tukuyu focus of southwest Tanzania after ten years of ivermectin mass treatment. Tanzan Health Res Bull. 2007; 9(3): 174-9.
332901	Tanzania	Makunde WH, Salum FM, Massaga JJ, Alilio MS. Clinical and parasitological aspects of itching caused by onchocerciasis in Morogoro, Tanzania. Ann Trop Med Parasitol. 2000; 94(8): 793–9.
327919	Togo	Golden A, Faulx D, Kalnoky M, Stevens E, Yokobe L, Peck R, Karabou P, Banla M, Rao R, Adade K, Gantin RG, Komlan K, Soboslay PT, de Los Santos T, Domingo GJ. Analysis of age-dependent trends in Ov16 IgG4 seroprevalence to onchocerciasis. Parasites Vectors. 2016; 9(1): 338.
338602	Togo	Dogba PK, Komi BT. [The evaluation of endemic onchocerciasis in the Amou prefecture (Togo)]. Bull Soc Pathol Exot. 1994; 87(2): 110–1.
327978	Uganda	Katabarwa M, Lakwo T, Habomugisha P, Agunyo S, Byamukama E, Oguttu D, Ndyomugyenyi R, Tukesiga E, Ochieng GO, Abwaimo F, Onapa A, Lwamafa DWK, Walsh F, Unnasch TR, Richards FO. Transmission of Onchocerca volvulus by Simulium neavei in Mount Elgon focus of Eastern Uganda has been interrupted. Am J Trop Med Hyg. 2014; 90(6): 1159-66.
328051	Uganda	Katabarwa MN, Lakwo T, Habomugisha P, Agunyo S, Byamukama E, Oguttu D, Tukesiga E, Unoba D, Dramuke P, Onapa A, Tukahebwa EM, Lwamafa D, Walsh F, Unnasch TR. Transmission of Onchocerca volvulus continues in Nyagak-Bondo focus of northwestern Uganda after 18 years of a single dose of annual treatment with ivermectin. Am J Trop Med Hyg. 2013; 89(2): 293-300.
332836	Uganda	Kipp W, Bamhuhiiga J, Rubaale T, Kabagambe G. Adverse reactions to the ivermectin treatment of onchocerciasis patients: does infection with the human immunodeficiency virus play a role?. Ann Trop Med Parasitol. 2005; 99(4): 395-402.

NID	Geographies	Citation
332845	Uganda	Ndyomugyenyi R, Tukesiga E, Büttner DW, Garms R. The impact of ivermectin treatment alone and when in parallel with Simulium neavei elimination on onchocerciasis in Uganda. Trop Med Int Health. 2004; 9(8): 882-6.
333505	Uganda	Fischer P, Kipp W, Bamuhiga J, Binta-Kahwa J, Kiefer A, Büttner DW. Parasitological and clinical characterization of Simulium neavei-transmitted onchocerciasis in western Uganda. Trop Med Parasitol. 1993; 44(4): 311–21.
333506	Uganda	Fischer P, Kipp W, Kabwa P, Buttner DW. Onchocerciasis and human immunodeficiency virus in western Uganda: prevalences and treatment with ivermectin. Am J Trop Med Hyg. 1995; 53(2): 171–8.
334413	Uganda	Katabarwa M, Onapa AW, Nakileza B. Rapid epidemiological mapping of onchocerciasis in areas of Uganda where Simulium neavei sl is the vector. East Af Med J. 1999; 76(8): 440–6.
334415	Uganda	Katabarwa MN, Walsh F, Habomugisha P, Lakwo TL, Agunyo S, Oguttu DW, Unnasch TR, Unoba D, Byamukama E, Tukesiga E, Ndyomugyenyi R, Richards FO. Transmission of Onchocerciasis in Wadelai Focus of Northwestern Uganda Has Been Interrupted and the Disease Eliminated [Internet]. J Parasitol Res. 2012.
334417	Uganda	Katabarwa M. Has interruption of Simulium neavei s.s transmitted onchocerciasis been attained in the Kashoya-Kitomi focus?. 2010.
334419	Uganda	Kipp W, Bamuhiga JT, Kwered EM. Onchocerciasis prevalence in previously known foci in western Uganda: results from a preliminary survey in Kabarole district. Trop Med Parasitol. 1992; 43(2): 80–2.
334420	Uganda	Lakwo TL, Garms R, Rubaale T, Katabarwa M, Walsh F, Habomugisha P, Oguttu D, Unnasch T, Namanya H, Tukesiga E, Katamanywa J, Bamuhiiga J, Byamukam E, Agunyo S, Richards F. The disappearance of onchocerciasis from the Itwara focus, western Uganda after elimination of the vector Simulium neavei and 19 years of annual ivermectin treatments. Acta Trop. 2013; 126(3): 218–21.
334422	Uganda	Okello DO, Ovuga EB, Ogwal-Okeng JW. Dermatological problems of onchocerciasis in Nebbi District. Uganda East Afr Med J. 1995: 72(5): 295–8



76 Supplementary Figure 3. Africa and Yemen data coverage maps.

177 Locations of the unique point and polygon data used in modelling, grouped by years: 1988–1999, 2000–2004, 2005–

178 2009, and 2010–2018. Hatched countries were not included in the MBG modelling region. Data that were able to be matched to point coordinates are represented by red dots and data that represented a larger area are represented by

180 blue polygons.

181 **3.5 APOC survey year matching**

182 The African Programme for Onchocerciasis Control (APOC),(3) which operated from 1995 to 2015, relied on Rapid 183 Epidemiological Mapping of Onchocerciasis (REMO) to map baseline onchocerciasis prevalence and endemicity in 184 19 onchocerciasis-endemic African countries outside the Onchocerciasis Control Programme (OCP) focal region 185 (APOC was later expanded to additional countries). REMO(4,5) mapping, involving village-level surveys of nodule 186 (onchocercoma) prevalence among adult males, was completed for 12 APOC countries by 2001.(6) We obtained 187 these pre-2001 nodule mapping data, when available, from the ESPEN data portal, although detailed information on 188 survey years were lacking for many (N = 4409) of these datapoints. So that these essential baseline mapping surveys 189 could inform our spatiotemporal model of onchocerciasis prevalence, we investigated the pre-2001 REMO mapping 190 surveys in APOC countries and attempted to narrow the possible range of survey years for these data. As widespread 191 use of REMO by APOC began in 1996,(6) and these data were indicated by the ESPEN database as representing 192 pre-2001 surveys, we considered 1996 and 2000 as the initial lower and upper bounds for survey years. We next 193 identified survey locations for which the first year of onchocerciasis MDA (mass drug administration) occurred prior 194 to 2001 (see section 4.2 below) and assumed that baseline mapping occurred at that location no later than the 195 preceding year, treating that year as the new upper bound. Finally, we examined the available literature on pre-2001 196 APOC REMO surveys, by country, to further narrow the possible range of survey years. Our decisions around 197 survey year estimates are summarised in Supplementary Table 4. For each datapoint, we specified the final survey 198 year as the median of the lower and upper year bounds (rounding down), considering this approach to balance the 199 potential conflicting biases from incorrect survey years on the temporal trends in the MBG model and the 200 association with environmental and sociodemographic covariates in child models.

201

202 Survey year data were also missing for some Phase 1a and 1b monitoring and evaluation surveys in the APOC data 203 retrieved from ESPEN. We identified survey years for most of these data by matching them to the locations, sample 204 sizes, cases, and number of villages reported by Tekle et al.,(7) supplemented with additional sources(8,9) for some 205 countries. We identified survey years for a total of 1001 observations. Survey years could not be reliably identified 206 for some Phase 1a and 1b data, specifically Phase 1a data for Kasai Occidental (Democratic Republic of the Congo, 207 DRC), Phase 1a and 1b data for Bioko Island (Equatorial Guinea), Phase 1a data for Kogi (Nigeria), and some Phase 208 1b data for Uganda; these data were excluded from our analysis. We assumed sample age ranges of 5-99 years of 209 age for Phase 1a and 1b data(7) unless the ESPEN dataset indicated otherwise.

210 Supplementary Table 4. APOC REMO survey year investigations. CDTI: Community-directed treatment with 211 ivermectin.

Country	Data Rows	Notes	Survey Year Strategy
Central African Republic	875	CDTI began in some areas in 1993, following REMO surveys.(10)	Use 1992 (one year prior to CDTI initiation) as lower bound.
Cameroon	389	Early proof-of-concept tests of the REMO methodology were performed in Cameroon in 1993.(4)	Assume 1993 as lower bound.
Democratic Republic of Congo	684	REMO began in 1997 in certain foci and continued until 2008;(11) CDTI areas were established in 1999 based on REMO.(12)	REMO appears to have been undertaken between 1997 and 1999 (although Bof(11) noted that it continued through 2008; ESPEN data include surveys beyond 2001). Assume 1997 as lower bound for pre-2001 DRC nodule surveys.
Republic of Congo	290	No information found except an MDA start year of 2001 for a small subset of these locations.	Assume 1996 as lower bound.
Ethiopia	283	Country-wide REMO was performed in Ethiopia in 1997 and 2001.(13)	Use 1997 as survey year for pre-2001 data.
Kenya	93	No information found.	Assume 1996 as lower bound for pre-2001 data.

Country	Data Rows	Notes	Survey Year Strategy
Nigeria	576	REMO was reported in 1995 to have been used for nationwide mapping;(3) another source indicated that REMO was carried out in Nigeria between 1994 and 1996.(14)	Assume 1994 and 1995 as lower and upper bounds of survey years for pre-2001 data.
Rwanda	90	Nationwide REMO was performed in 1999.(15)	Use 1999 as survey year for pre-2001 data.
Sudan	2	No information found.	Assume 1996 as lower bound.
South Sudan	110	REMO was performed between 1995 and 2002.(16)	Assume 1995 as lower bound.
Chad	481	REMO performed prior to 1999.(17)	Assume 1996 as lower bound, and 1998 as upper bound for surveys otherwise lacking upper bound estimates based on MDA.
Tanzania	142	Years of initiation of CDTI in Tanzania ranged from 1995 to 2004.(7)	Use 1994 as lower bound estimate.
Uganda	394	Nationwide REMO conducted between 1993 and 1997.(18)	Use 1993 and 1997 as lower and upper bounds, respectively.

213 4.0 Supplementary covariates

214 4.1 Pre-existing covariates considered for analysis

215 A variety of environmental and sociodemographic variables were used to predict all-age onchocerciasis prevalence.

216 Where available, the finest spatiotemporal resolution of gridded datasets was used. Geospatial covariate rasters were

217 resampled to ~ 5 km in GeoTiff format, for consistent modelling. Where data coverage was inconsistent between our

standard mask and the input data, either a local average or nearest neighbour method (depending on data type) was

219 used to fill spatial data gaps. Data from the nearest year available were used if covariate coverage did not include all

220 model years. Supplementary Table 5 contains a full list of covariates considered in our analysis.

221 4.2 Creation of MDA covariate

222 Data used in the creation of the onchocerciasis MDA covariate were downloaded from the ESPEN Portal in April 223 2019.(19) The ESPEN data included the MDA start year and cumulative number of MDA rounds for each IU. 224 ArcGIS software was then used to join the MDA data to the NTD (Neglected Tropical Disease) Implementation 225 Unit Shapefile maintained by the Task Force for Global Health. A custom polygon was created for Sudan by 226 referencing annual reports from The Carter Center. The MDA covariate value was set calculated as the cumulative 227 number of rounds (defined as total number of rounds implemented) for the IU, and then converted to a raster for use 228 in geospatial analysis. Data for lymphatic filariasis MDA were obtained from WHO(20) for the years during which 229 LF (lymphatic filariasis) MDA was conducted, by IU, and joined onto the IU shapefile maintained by ESPEN. So 230 that the effects of MDA for onchocerciasis and lymphatic filariasis were not double-counted in locations where both 231 were indicated, a composite MDA covariate was produced which indicated the cumulative number of years for 232 which either onchocerciasis or lymphatic filariasis MDA was indicated. This composite covariate was used in 233 modelling.

234

235 **4.3** Covariate reduction

High collinearity among covariates may lead to unstable model coefficients and unreliable predictions.³⁷ To reduce

this problem, we derived a reduced covariate set using analysis of variance inflation factors (VIF).(22) Starting with

- the full list of covariates, we iteratively removed covariates with the highest VIF values until all remaining
- covariates had a VIF below 3.0. The reduced covariate set was used in fitting the MBG model and for
- spatiotemporal predictions. Supplementary Table 5 indicates the covariates that were retained in the final model,
- with representative plots provided for each covariate in Supplementary Figure 4. All variables were matched to the

- 243 year of reported prevalence data, without any temporal lags (eg, temperature values for the year 2000 were joined to prevalence data for the year 2000).

244 Supplementary Table 5. Covariates considered or retained for modelling, 1988–2018.

245 All covariates considered for inclusion as predictors of onchocerciasis prevalence. Short names are provided for reference in later figures and tables. Covariates

246 247 retained after VIF analysis are indicated with a (+).

Covariate	Short Name	Retained in Final Model	Years Available	Source	Reference
Aridity	crutsard	-	1988–2018	Climatic Research Unit Time-Series (CRUTS)	Harris, I., Osborn, T.J., Jones, P. et al. Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset. Sci Data 7, 109 (2020). <u>https://doi.org/10.1038/s41597-020-0453-3</u>
					University of East Anglia. Climatic Research Unit TS v. 4.04 dataset. Available at: https://crudata.uea.ac.uk/cru/data/hrg/
Average daily mean temperature	crutstmp	+	1988–2018	CRUTS	Harris, I., Osborn, T.J., Jones, P. et al. Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset. Sci Data 7, 109 (2020). <u>https://doi.org/10.1038/s41597-020-0453-3</u>
					University of East Anglia. Climatic Research Unit TS v. 4.04 dataset. Available at: https://crudata.uea.ac.uk/cru/data/hrg/
Average daily minimum temperature	crutstmn	-	1988–2018	CRUTS	Harris, I., Osborn, T.J., Jones, P. et al. Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset. Sci Data 7, 109 (2020). <u>https://doi.org/10.1038/s41597-020-0453-3</u>
					University of East Anglia. Climatic Research Unit TS v. 4.04 dataset. Available at: https://crudata.uea.ac.uk/cru/data/hrg/
Average daily maximum temperature	crutstmx	-	1988–2018	CRUTS	Harris, I., Osborn, T.J., Jones, P. et al. Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset. Sci Data 7, 109 (2020). <u>https://doi.org/10.1038/s41597-020-0453-3</u>
					University of East Anglia. Climatic Research Unit TS v. 4.04 dataset. Available at: https://crudata.uea.ac.uk/cru/data/hrg/
Diurnal temperature range	crutsdtr	+	1988–2018	CRUTS	Harris, I., Osborn, T.J., Jones, P. et al. Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset. Sci Data 7, 109 (2020). <u>https://doi.org/10.1038/s41597-020-0453-3</u>
					University of East Anglia. Climatic Research Unit TS v. 4.04 dataset. Available at: https://crudata.uea.ac.uk/cru/data/hrg/

Covariate	Short Name	Retained in Final Model	Years Available	Source	Reference
Vapor pressure	crutsvap	-	1988–2018	CRUTS	Harris, I., Osborn, T.J., Jones, P. et al. Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset. Sci Data 7, 109 (2020). <u>https://doi.org/10.1038/s41597-020-0453-3</u>
					University of East Anglia. Climatic Research Unit TS v. 4.04 dataset. Available at: https://crudata.uea.ac.uk/cru/data/hrg/
Wet day frequency	crutswet	+	1988–2018	CRUTS	Harris, I., Osborn, T.J., Jones, P. et al. Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset. Sci Data 7, 109 (2020). <u>https://doi.org/10.1038/s41597-020-0453-3</u>
					University of East Anglia. Climatic Research Unit TS v. 4.04 dataset. Available at: https://crudata.uea.ac.uk/cru/data/hrg/
Distance to rivers >25 m wide	distrivers25m	+	Synoptic	Natural Earth Data (derived)	Andreadis KM, Schumann GJ-P, Pavelsky T. A simple global river bankfull width and depth database. Water Resources Research. 2013;49(10):7164–8.
River width (largest river within 5x5 km cell)	river_size	+	Synoptic	Natural Earth Data (derived)	Andreadis KM, Schumann GJ-P, Pavelsky T. A simple global river bankfull width and depth database. Water Resources Research. 2013;49(10):7164–8.
Elevation	elevation	-	Synoptic	NOAA/NCEI	Young, A. H., K. R. Knapp, A. Inamdar, W. B. Rossow, and W. Hankins, 2017: "The International Satellite Cloud Climatology Project, H-Series Climate Data Record Product", Earth System Science Data, in preparation.
Enhanced Vegetation Index (EVI)	evi_v6	+	2000–2018	MODIS	Huete, A., Justice, C. & van Leeuwen, W. MODIS vegetation index (MOD 13) algorithm theoretical basis document. (1999).
					USGS & NASA. Vegetation indices 16-Day L3 global 500m MOD13A1 dataset. Available at: https://lpdaac.usgs.gov/dataset_discovery/modis/m odis_products_table/mod13a1. (Accessed: 25th July 2017)
					Weiss, D. J. et al. An effective approach for gapfilling continental scale remotely sensed timeseries. Isprs J. Photogramm. Remote Sens. 98, 106–118 (2014).
					C. Schaaf, Z. Wang. (2015). MCD43A1 MODIS/Terra+Aqua BRDF/Albedo Model Parameters Daily L3 Global - 500m V006. NASA EOSDIS Land Processes DAAC. http://doi.org/10.5067/MODIS/MCD43A1.006
Population	worldpop	+	1988–2018	WorldPop	Lloyd, C. T., Sorichetta, A. & Tatem, A. J. High resolution global gridded data for use in population studies. Sci. Data 4, sdata20171 (2017).
					World Pop. Get data. Available at: <u>http://www.worldpop.org.uk/data/get_data/</u> . (Accessed: 25th July 2017)

Covariate	Short Name	Retained in Final Model	Years Available	Source	Reference
Growing season length	growingseason	-	Synoptic	FAO	FAO. GAEZ - Global Agro-Ecological Zones data portal. Available at: http://www.fao.org/nr/gaez/about-data-portal/en/. (Accessed: 25th July 2017)
					FAO. GAEZ - Global Agro-Ecological Zones users guide. (2012).
Tassled cap brightness	tcb_v6	+	2000–2017	MODIS	USGS & NASA. Nadir BRDF- Adjusted Reflectance Reflectance 16-Day L3 Global 1km dataset. Available at: https://lpdaac.usgs.gov/dataset_discovery/modis/m odis_products_table/mcd43b4. (Accessed: 25th July 2017)
					 Strahler, A. H. & Muller, JP. MODIS BRDF/Albedo product: algorithm theoretical basis document version 5.0. (1999). Weiss, D. J. et al. An effective approach for gapfilling continental scale remotely sensed timeseries. Isprs J. Photogramm. Remote Sens. 98, 106–118 (2014).
					C. Schaaf, Z. Wang. (2015). MCD43A1 MODIS/Terra+Aqua BRDF/Albedo Model Parameters Daily L3 Global - 500m V006. NASA EOSDIS Land Processes DAAC. http://doi.org/10.5067/MODIS/MCD43A1.006
Precipitation (Multi- source Weighted- Ensemble)	mswep	+	1988–2016	Princeton Climate Analytics	Beck, H.E., A.I.J.M. van Dijk, V. Levizzani, J. Schellekens, D.G. Miralles, B. Martens, A. de Roo: MSWEP: 3-hourly 0.25 global gridded precipitation (1979-2015) by merging gauge, satellite, and reanalysis data, Hydrology and Earth System Sciences, 21(1), 589-615, 2017.
Slope for land surfaces	slope	+	Synoptic	NOAA/NCEI	Young, A. H., K. R. Knapp, A. Inamdar, W. B. Rossow, and W. Hankins, 2017: "The International Satellite Cloud Climatology Project, H-Series Climate Data Record Product", Earth System Science Data, in preparation.
Soil: Saturated water content (200 cm depth)	sgawets	-	Synoptic	SoilGrid	Hengl T, Mendesde Jesus J, MacMillan RA, Batjes NH, Heuvelink GBM, Ribeiro E, Samuel- Rosa A, Kempen B, Leenaars JGB, Walsh MG, Gonzalez MR. "SoilGrids1km — Global soil information based on automated mapping." <i>PLOS ONE</i> 9(8): e105992. 29 Aug 2014. doi:10.1371/journal.pone.0105992
Soil: Probability of bedrock exposure	sgbdrlog	+	Synoptic	SoilGrid	Hengl T, Mendesde Jesus J, MacMillan RA, Batjes NH, Heuvelink GBM, Ribeiro E, Samuel- Rosa A, Kempen B, Leenaars JGB, Walsh MG, Gonzalez MR. "SoilGrids1km — Global soil information based on automated mapping." <i>PLOS ONE</i> 9(8): e105992. 29 Aug 2014. doi:10.1371/journal.pone.0105992
Soil: Bulk density (200 cm depth)	sgbldfie	+	Synoptic	SoilGrid	Hengl T, Mendesde Jesus J, MacMillan RA, Batjes NH, Heuvelink GBM, Ribeiro E, Samuel- Rosa A, Kempen B, Leenaars JGB, Walsh MG, Gonzalez MR. "SoilGrids1km — Global soil information based on automated mapping." <i>PLOS ONE</i> 9(8): e105992. 29 Aug 2014. doi:10.1371/journal.pone.0105992
Soil: Clay content mass fraction (200 cm depth)	sgclyppt	+	Synoptic	SoilGrid	Hengl T, Mendesde Jesus J, MacMillan RA, Batjes NH, Heuvelink GBM, Ribeiro E, Samuel- Rosa A, Kempen B, Leenaars JGB, Walsh MG, Gonzalez MR. "SoilGrids1km — Global soil

Covariate	Short Name	Retained in Final Model	Years Available	Source	Reference
					information based on automated mapping." <i>PLOS ONE</i> 9(8): e105992. 29 Aug 2014. doi:10.1371/journal.pone.0105992
Soil: Coarse fragments (200 cm depth)	sgcrfvol	+	Synoptic	SoilGrid	Hengl T, Mendesde Jesus J, MacMillan RA, Batjes NH, Heuvelink GBM, Ribeiro E, Samuel- Rosa A, Kempen B, Leenaars JGB, Walsh MG, Gonzalez MR. "SoilGrids1km — Global soil information based on automated mapping." <i>PLOS ONE</i> 9(8): e105992. 29 Aug 2014. doi:10.1371/journal.pone.0105992
Cumulative years of mass drug administration (MDA) for lymphatic	allmda	+	1988–2018	WHO (rasters produced in current study)	WHO. Expanded Special Project for Elimination of Neglected Tropical Diseases (ESPEN). https://www.espen.org/ (accessed May 20, 2020).
filariasis or onchocerciasis					WHO. Global Programme to Eliminate Lymphatic Filariasis. http://www.who.int/lymphatic_filariasis/elimination- programme/en/ (accessed May 29, 2019).



251 Supplementary Figure 4. Africa and Yemen covariate values.

15 environmental or sociodemographic variables were used as inputs for the modelling process for Africa and
 Yemen (river size is not shown in this figure). Time-varying covariates are presented here for the year 2015. Please

254 refer to Supplementary Table 5 for the corresponding citations for each covariate.

255 5.0 Supplementary methods

256 5.1 Age and diagnostic crosswalks

- 257 Surveys of onchocerciasis prevalence have varied in both their sampled age ranges and in the diagnostic tests they
- 258 utilised. For example, epidemiological surveys during the OCP program relied on skin snip (microfiladermia)
- 259 examinations of individuals aged 5 years or older,(23,24) while APOC relied on the REMO (Rapid Epidemiological
- 260 Mapping of Onchocerciasis)(5,6,25,26) methodology to identify endemic areas requiring MDA, employing nodule
- 261 palpation in adults. To harmonise these data sources, we adjusted survey data to represent all-age microfiladermia

- 262 prevalence by developing and applying an age and diagnostic crosswalk model that simultaneously accounts for
- typical age-dependent trends in prevalence and differential detection sensitivity between skin snip examination and
 nodule palpation. We did not crosswalk results from antibody (eg, Ov16) surveys as these data were not used in
 fitting the MBG model.
- 265 fitting the MBG mod 266

267 We identified published within-study comparisons reporting results from skin snip biopsies (microfiladermia), 268 nodule (onchocercoma) palpation, or both, for more than one age group in a given study population. Study cohorts 269 with zero reported cases were excluded from the crosswalk training dataset because they do not contribute 270 information with which to derive age trends in endemic settings. Studies were eligible for inclusion in the crosswalk 271 training dataset if they reported data from a country in our MBG modelling region; all eligible studies reported data 272 from surveys conducted in our geospatial modelling timeframe (1988–2018). We identified 133 unique survey 273 populations from 36 studies reporting skin snip prevalence in multiple age groups, and 126 unique survey 274 populations from 22 studies reporting nodule prevalence in multiple age groups; among these surveys, a total of 100 275 unique survey populations from 19 studies reported both skin snip and nodule prevalence from the same study 276 populations and age groups. Supplementary Table 6 summarises these sources.

277 Supplementary Table 6. Data used in estimation of age and diagnostic crosswalk.

Note that studies may have reported data for additional diagnostic tests that were not used in crosswalk model
 fitting.

Countries	Survey Years	Diagnostic Data
Benin(27)	1991	skin snip, nodule
Cameroon(28)	1992	skin snip
Cameroon(29)	1992	skin snip, nodule
Cameroon(30)	1996, 2005, 2006, 2011	skin snip, nodule
Cameroon(31)	2000	nodule
Cameroon(32)	2008	skin snip, nodule
Cameroon(33)	2009	skin snip, nodule
Cameroon(34)	2009	skin snip, nodule
Cameroon(35)	2012	skin snip, nodule
Cameroon(36)	2015	skin snip, nodule
Cameroon(37)	2015	skin snip, nodule
Cameroon(38)	2015	skin snip, nodule
Cameroon, Uganda(39)	1993 (Uganda), 1996 (Cameroon), 2005 (both countries)	skin snip, nodule
Central African Republic(40)	1990, 1995	skin snip
Ethiopia(41)	1994	skin snip, nodule
Ethiopia(42)	1997	skin snip

Ethiopia(43)	2005	skin snip
Ethiopia(44)	2006	skin snip
Ethiopia(45)	2012	skin snip
Gabon(46)	2004	nodule
Nigeria(47)	1992	skin snip
Nigeria(48)	1994	skin snip, nodule
Nigeria(49)	1994	skin snip
Nigeria(50)	1997	skin snip
Nigeria(51)	2000	skin snip, nodule
Nigeria(52)	2005	skin snip
Nigeria(53)	2007	skin snip
Nigeria(54)	2008	skin snip, nodule
Nigeria(55)	2008	skin snip
Nigeria(56)	2008	skin snip, nodule
Nigeria(57)	2009	skin snip
Nigeria(58)	2009	skin snip, nodule
Nigeria(59)	2010	nodule
Sierra Leone (60)	2010	skin snip
Togo(61)	1992	skin snip
Togo(62)	2014	skin snip
Togo(63)	2015	skin snip
Uganda(64)	1993	skin snip, nodule
Uganda(65)	2012	skin snip, nodule

We first retrieved population estimates by single age-year from the Global Burden of Disease (GBD)(66) for the country and year of each survey in the crosswalk training set. We assumed that the age distribution (P(A), or probability of age A) within a survey sample matched that in the country and year of the survey, and estimated prevalence-by-age models (P(D|A), or the probability of disease, D, at age A) from birth through age 94 years, the maximum individual age-year modelled by GBD. We used the GBD age distributions to estimate the likelihood of observing the reported number of cases in each surveyed age bin, given a logistic (binomial) regression model of the average prevalence-by-age relationship across surveys:

288 289

290

 $logit(P(D|A)) = \beta_0 + I_{ss}f_{ss}(A) + I_{nod}f_{nod}(A) + \alpha_i + I_{ss}\beta_{ss}$

291 Onchocerciasis prevalence at a given age (P(D|A)) was modelled in logit space as a linear combination of an

292 intercept, β_0 , which was set at logit(0.00001) to drive prevalence at birth toward zero; basis splines (fda R

293 package(67)) on age, f(A), to accommodate non-linear age trends; α_i , cohort-level fixed effects (indicator variables

- 294 identifying study cohorts in the training dataset) to account for differences in study populations and survey designs;
- and a fixed diagnostic effect for skin snip surveys, β_{ss} , to model overall differences between skin snip and nodule
- surveys. Diagnostic indicator variables, I_{ss} and I_{nod} , were set to 1 for skin snip and nodule observations, respectively, and set to 0 otherwise, to select the appropriate age and diagnostic effects for a given survey. Study
- respectively, and set to 0 otherwise, to select the appropriate age and diagnostic effects for a given survey. Study cohort effects were assumed to be shared across diagnostic tests. The likelihood function was evaluated as the sum,
- 299 across studies (*i*) and age bins ($A1 \le A < A2$), of the binomial likelihood of observing the reported numbers of
 - across studies (*i*) and age onis ($A_1 \ge A < A_2$), of the officinial fixelihood of observing the reported numbers of cases given the reported sample sizes, age ranges, and diagnostic tests, using population-weighted mean prevalence values evaluated at the midpoints of each age year.
 - 302

Spline knot placements were identified by spacing four internal knots evenly by quantile in the training data, determined separately for skin snip and nodule surveys, with additional knots placed at ages 3, 6, and 65 to help stabilise model behaviour in early childhood and older adulthood. Starting values for all spline, cohort and diagnostic coefficients were randomly drawn from uniform distributions in the interval [-5, 5]. Models were estimated with maximum likelihood optimisation using the box constraint quasi-Newton algorithm⁶⁶ (without constraints) in the optim function (R stats package(69)).

- 309
- Prior to running the MBG model, all survey data that derived from a restricted age range (ie, anything other than 0– 94 years), or that represented nodule prevalence, were adjusted using the fitted crosswalk model. For each individual survey population, the crosswalk model was refit via maximum likelihood to the prevalence data for that survey, with all coefficients other than the cohort-level effect fixed, α_i , fixed to their estimates from the full crosswalk
- 314 model. This newly estimated value of α_i was then re-inserted into the crosswalk model with the other coefficients
- 315 fixed, to calculate all-age skin snip prevalence for that survey sample. Crosswalk model with the other coefficients
- 316 outcome measure in the MBG model.
- 317

Surveys that reported prevalence of 0% or 100% were particularly difficult to interpret for crosswalk purposes. A report of 0% prevalence could reflect true absence of infection, insufficient diagnostic sensitivity, or sampling variance in small samples, while a report of 100% prevalence masks any implicit linkage between prevalence and the scale of infection intensity. While many of these concerns also apply to surveys reporting intermediate prevalence, the particular uncertainty involved with surveys reporting 0% or 100% prevalence increases the risk of inappropriate crosswalking, and for this reason we did not crosswalk such surveys.

Uncertainty in the crosswalk models was estimated using bootstrap analysis, with 100 replicates generated by sampling, with replacement, an equal number of study cohorts as in the full crosswalk training dataset. Resampling

was conducted by cohort rather than age bin to better estimate variation among surveys. Spline knot locations and

328 model coefficients were estimated for each bootstrap replicate independently of other replicates and of the full 329 model. To visualise bootstrapped results for a given survey, cohort effects were calculated for each replicate using

maximum likelihood optimisation with all other coefficients fixed to the estimated values for the replicate.





334 335 Outputs from the diagnostic and age crosswalk model. (a) Example prevalence-by-age curves fit to a study from Benin(27) that reported both skin snip and nodule age-binned data and was included in the crosswalk model training

336 dataset. Estimates are shown for skin snip microfiladermia prevalence (solid black line) and nodule prevalence

337 (dotted green line); horizontal bars indicate the age-binned prevalence data reported by that study (solid gray: skin 338

snip; dotted green: nodule). (b) Bootstraped estimates of differences between microfiladermia (skin snip) and nodule

339 prevalence by age, in logit space. Each of 100 bootstrap samples is shown (faint grey line) along with the median 340 estimate (solid black line) and central 95% uncertainty interval (shaded area). Positive values indicate higher skin

341 snip prevalence than nodule prevalence.

343 In an example location with moderately high microfiladermia prevalence, our final crosswalk model (Supplementary 344 Figure 5) suggests a sharp increase in prevalence through childhood and adolescence, with a slower increase 345 between approximately ages 15 and 65 years. This general pattern is qualitatively consistent with the range of 346 onchocerciasis microfiladermia age trends reported by other studies (70-72) although substantial site-specific 347 variation exists. The relationship between microfiladermia and nodule prevalence within onchocerciasis-endemic 348 communities was previously modelled by Coffeng et al.,(73) using pre-control data from a broad sampling of sites in 349 OCP and APOC regions. Their multivariate logistic regression model, relating nodule prevalence in adult males 350 (aged 20 years or older) to microfiladermia prevalence in individuals aged 5 years or older, did not explicitly model 351 changes in prevalence with age, but revealed higher prevalence estimates from skin snip biopsies than those from 352 nodule palpation surveys in the same communities. Our full crosswalk model and bootstrapped uncertainty estimates 353 (Supplementary Figure 5) are qualitatively consistent with their results, estimating that skin snip prevalence exceeds

nodule prevalence, on average, from adolescence until about age 50, with poorer resolution of this relationship in

354 355 356

357 While crosswalking survey data that are based on differing diagnostics and age coverage enabled us to leverage a 358 more comprehensive geospatial dataset than is otherwise tractable, we identify several limitations to our crosswalk 359 approach. Our model assumes that, for a given diagnostic approach, changes in prevalence by age follow a 360 consistent pattern across locations, years, sexes, and programmatic contexts. However, actual and reported 361 microfiladermia and nodule prevalence patterns are influenced by local factors including ecological conditions; 362 infection intensity; vector identity, density, exposure, and control history; MDA coverage; variations in survey 363 sampling designs and in diagnostic specificity and sensitivity; and sex.(70,72–79) We have not modelled the age-364 and diagnostic-specific effects of MDA on prevalence in the crosswalk models (although MDA is included as a 365 predictor during the MBG modelling stage). We were also limited by the variable reporting of age information 366 among data sources. Some surveys did not report age ranges, and for these surveys we assumed that their data 367 represented all-age prevalence, risking possible misclassification. The absence of individual-level data on onchocerciasis prevalence also precluded full age-standardisation, as we could only assume that the survey sample 368 369 matched the age structure of the general population. Our crosswalks do not currently account for the sensitivity and 370 specificity of skin snip and nodule diagnostics, and crosswalk uncertainty is therefore likely to be underestimated. 371 We also do not currently have a computationally feasible method to propagate uncertainty from the crosswalk 372 models into the MBG models in a way that accounts for the inconsistent reporting of sampled age ranges among 373 studies.

374 375

376 5.2 Polygon resampling

children and older adults.

377 Prevalence records are representative of either georeferenced point locations or polygonal areas (eg, as defined by 378 the borders of administrative or programmatic units). As our modelling framework relies on coordinate-referenced 379 data in order to fit the continuous spatial random fields, we converted areal data to a representative collection of 380 point data. This "polygon resampling" process, described previously for geostpatial modelling of under-5 381 mortality,(80) generates candidate locations based on the underlying population density of the resampled area, 382 implicitly assuming that surveys employed population-based designs, and is illustrated in Supplementary Figure 6 383 using Cameroon in 2015 as an example. For each polygonal observation in our dataset, 10 000 points were randomly 384 sampled from within the polygon, with weighting by the WorldPop total population raster. Candidate points were 385 clustered using k-means clustering, generating a set of final points with a density of 1 per 1000 grid cells, except for 386 small polygons, in which case density was iteratively increased by a factor of 10 until a minimum threshold of 10 387 points was achieved. Weights were assigned to each point proportionally to the number of candidate points that 388 entered into the k-means cluster. The points generated by this resampling process were assigned the prevalence of 389 onchocerciasis reported for the survey for that polygon. These sample weights were used in MBG model fitting

390 within INLA.





392 393 S

3 Supplementary Figure 6. Polygon resampling.

The process of polygon resampling is illustrated using reported onchocerciasis prevalence data for Cameroon in 2015. (a) National map of Cameroon, showing the inset area (red frame) that is featured in the remaining panels. (b) Reported areal (irregular polygons) and point-level (circles) prevalence data. (c) Underlying population surface from WorldPop (displayed on a log₁₀ scale), with survey polygons overlayed. Polygons are resampled proportionally to 5 x 5 km population density. (d) Locations of the final datapoints for geospatial modelling, showing both those data originally reported with coordinates and those derived by resampling the polygon data. The opacity of resampled datapoints in (d) reflects their relative weights, which sum to 1.0 within an individual areal survey.

401 5.3 Geostatistical model

402 5.3.1 Model geographies and time period

403 Model-based geostatistical (MBG) methods were used to generate estimates of all-age onchocerciasis

404 microfiladermia prevalence for onchocerciasis-endemic countries of Africa, plus Yemen (listed in Supplementary

- 405 Table 2). A single model was fit to this geographical region. We did not model countries in the Western Hemisphere
- 406 that were formerly or are currently endemic for onchocerciasis, due to their highly localised onchocerciasis foci. We
- 407 were principally concerned with estimates for the time period 2000–2018, but fit the model using data from 1988–
- 408 2018 in order to leverage information from pre-2000 OCP and APOC surveys, and thereby improve "baseline" (ie,
- 409 year 2000) model estimates in countries covered by those programmes. Reporting of results in the main text focuses
- 410 411 on estimates for 2000–2018.

412 5.3.2 Covariate coverage

413 As with any regression model, the reliability of predictions from our model is affected by the overlap between

- 414 covariate values in training and prediction datasets. Predictions in regions with a range of covariate values that fall
- 415 outside the range of values in the training set may be prone to extrapolation errors. Supplementary Figure 7
- 416 illustrates the number of covariates whose mean values (averaged over 1988–2018) fall outside the central 95%
- 417 quantile interval of values in the training set, for each 5 x 5-km pixel. Child model predictions in areas with poor 418
- covariate representation (ie, a large number of covariates with values outside the central interval) should be 419
- considered with special caution. These areas include the Sahel and Sahara, as well as Yemen, Kenya, Somalia,
- 420 eastern Ethiopia, and southern Angola. 421

422 5.3.3 Environmental suitability

423 Cromwell et al. recently produced an environmental suitability model for onchocerciasis, using a boosted regression 424 tree (BRT) model trained with space- and time-referenced data on onchocerciasis occurrence. (81) In contrast to our

- 425 present MBG model, the Cromwell suitability model leveraged data from not only skin snip and nodule palpation
- 426 surveys, but also Ov16 serosurveys, onchocerciasis-derived eye or skin morbidity surveys, and other diagnostics.
- 427 The model was trained using an overlapping but non-identical set of environmental covariates as those used in the
- 428 present study, including climatic, topographic, hydrologic, vegetative, and urbanicity variables. Pseudo-absence 429
- records were generated using background sampling. Importantly for our present purposes, this suitability model does 430 not incorporate the influence of programmatic interventions for onchocerciasis and does not use data on
- 431 onchocerciasis absence, providing a reflection of O. volvulus endemicity apart from temporal changes in infection
- 432 prevalence. The outputs of the BRT model represent a spatially explicit index from 0–1, reflecting environmental
- 433 suitability for onchocerciasis occurrence in a given 5 x 5-km cell. We incorporated mean predictions from this
- 434 suitability model, using covariate values for 2016 (Supplementary Figure 8), to improve the behaviour of our MBG
- 435 model in areas of poor prevalence data coverage, particularly in areas where covariates lie outside the range of
- 436 values in sampled locations and MBG predictions are therefore subject to extrapolation uncertainty.



437 438 439 Supplementary Figure 7. Number of covariates with values outside the central 95% interval of values at survey sites.

440 For each 5 x 5-km pixel, the number of covariates (out of a total of 16) whose average value lies outside the central 441 95% interval of values at observed survey sites is indicated. Localities with a large number of such covariates

- 442 represent areas in which predictions are less reliable, due to potential extrapolation error.
- 443 444 445





448 Supplementary Figure 8. Onchocerciasis suitability model predictions (Cromwell et al., 2021) for 2016.

449 Mean environmental suitability for onchocerciasis, from a previous boosted regression tree (BRT) analysis (81) of
 450 onchocerciasis presence data, using covariate values for 2016.

452 **5.3.4 Model description**

We modelled *Onchocerca volvulus* infection prevalence using a spatially explicit Bayesian generalised linear mixed
 effects regression model in R-INLA:

456 $Y_{i,t} \sim \text{Binomial}(p_{i,t}, N_{i,t})$

458
$$\operatorname{logit}(p_{i,t}) = \beta_0 + \beta X_{i,t} + \gamma_{c[i]} + U_i + Z_i + \epsilon_{i,t}$$

- $\begin{array}{l} 459\\ 460 \end{array} \qquad \qquad \qquad Z_i \sim \mathrm{GP}(0, \Sigma_{space}) \end{array}$
- 461

462
$$\epsilon_{i,t} \sim N(0, \sigma_{nug}^2)$$

- 463
- 464 The number of infected individuals $(Y_{i,t})$ among a sample $(N_{i,t})$ in location *i* and year *t* was modelled as a binomial
- 465 variable. This model specifies logit-transformed infection prevalence $(p_{i,t})$ as a linear combination of an intercept
- 466 for the modelling region (β_0); covariate fixed effects (coefficients, β , and values, $X_{i,t}$); country random effects
- 467 $(\gamma_{c[i]})$; a second-order random walk model on estimates of onchocerciasis environmental suitability (U_i) ; a spatially
- 468 correlated random field (Z_i); and an uncorrelated error term or nugget effect ($\epsilon_{i,t}$). The spatial random field (Z_i) was
- 469 modelled as a Gaussian process with mean 0 and a Matérn covariance function (Σ_{space}). Modelled outputs from the
- 470 BRT analysis of onchocerciasis environmental suitability (U_i) , described in section 5.3.3, were modelled with a 471 second-order random walk, with values grouped into 25 bins by quantile. This random walk model accommodates
- 471 second-order random walk, with values grouped into 25 bins by quantile. This random walk model accommodates 472 non-linearity in the relationship between suitability (based on presence-absence data) and the linear predictor. The
- 473 INLA (integrated nested Laplace approximation) model was fit using an "empirical Bayes" integration strategy,
- 474 which relies on mode values to approximate hyperparameter posterior distributions during estimation and enabled us
- 475 to achieve tractable model computation times.(82)

477 **5.3.5 Priors**

- 478 We specified minimally informative priors for INLA hyperparameters, as detailed in Supplementary Table 7. Priors
- 479 for the spatial hyperparameters τ and κ were derived automatically by R-INLA based on the finite elements mesh.

480 Supplementary Table 7. INLA model priors.

Parameter	Description	Prior
β_0, β	Intercept, covariate fixed effects	$N(\mu = 0, \sigma^2 = 3^2)$
$\left(\frac{1}{\sigma_{country}^2}\right)$	Precision for country random effects ($\gamma_{c[i]}$, i.i.d.)	gamma(a = 1, b = 0.00005)
$\left(rac{1}{\sigma_{nug}^2} ight)$	Precision for nugget effect ($\epsilon_{i,t}$, i.i.d)	pc. prec $(u = 0.5, \alpha = 0.05)^1$
$\left(\frac{1}{\sigma_U^2}\right)$	Precision for suitability model (U_i , RW2 model)	gamma $(a = 1, b = 0.00005)$
σ_{SPDE}	Standard error for SPDE model	pc(3.0, 0.05) ²
Range	Range for SPDE model	pc(0.06171913, 0.05) ³

¹PC prior for precision.

²PC prior for SPDE σ , indicating a 5% probability that σ is greater than 3.

³PC prior for SPDE range, indicating a 5% probability that range is less than 0.06161412 (5% of the maximum extent of the spatial mesh).

481

482 5.3.6 Mesh construction

483 We modelled continuous spatial random effects using stochastic partial differential equations (SPDE)

484 representations of Gaussian-Markov random field (GMRF) approximations of a spatially autocorrelated Gaussian

485 process, using triangular finite element meshes as implemented in the R-INLA R package.(83–85) Due to the large

486 geographical size of the model region, a spherical (S2) mesh was constructed in order to minimise distance

487 distortions. Minimum and maximum edge lengths were set to 25 and 500 km, respectively, and a 1000-km external

488 buffer was used to avoid artifacts at the edges of the spatial domain.(84) These values were chosen to yield denser

489 mesh vertices in data-rich areas while maximising the overall spatial field resolution, within computational

490 constraints. The spatial mesh is illustrated in Supplementary Figure 9.



493 Supplementary Figure 9. Spatial mesh construction.

Two-dimensional projection of spherical refined Delaunay triangulation mesh used in estimating spatial random
 fields in Africa and Yemen, with national boundaries (bold lines). The mesh features greater vertex density in data rich locations.

497

498 5.3.7 Model fitting and estimation generation

499 Models were fit using the integrated nested Laplace approximation (INLA) algorithm in R-INLA. Fitted models for 500 each region were used to generate 1000 random samples from the joint posterior distributions of model parameters, 501 yielding mean and uncertainty estimates for onchocerciasis prevalence.

502

503 5.3.8 Model results

- 504 Model parameter estimates from the MBG model are summarised in Supplementary Table 8. Nominal range is the
- 505 distance (in km) at which spatial correlation has declined to about 0.1. Estimated random effects for the
- 506 environmental suitability covariate are displayed in Supplementary Figure 10.

507 Supplementary Table 8. Parameter estimates from in-sample onchocerciasis MBG model.

508	GP: Gaussian process. i.i.d.: Independent and identically distributed random effects. RW2: Second-order random
509	walk model.

Parameter	Median (95% UI)
Intercept	-3.656 (-4.214, -3.099)
GP nominal range	189.6 (176.7, 207.1)
GP nominal variance	3.177 (2.892, 3.568)
Nugget precision (i.i.d.)	0.719 (0.683, 0.742)
Country precision (i.i.d.)	0.505 (0.268, 0.992)
Environmental suitability precision (RW2)	32.269 (5.731, 125.009)
Covariate coefficients	
MDA (allmda)	-1.106 (-1.136, -1.077)
Diurnal temperature range (crutsdtr)	-0.363 (-0.537, -0.199)
Temperature (crutstmp)	-0.172 (-0.325, -0.018)
Wet day frequency (crutswet)	-0.163 (-0.315, -0.012)
Distance to rivers >25 m wide (distrivers25m)	-0.021 (-0.070, 0.028)
Enhanced Vegetation Index (evi_v6)	0.192 (0.129, 0.256)
Precipitation (mswep)	0.137 (0.057, 0.217)
Soil: Probability of bedrock exposure (sgbdrlog)	-0.063 (-0.119, -0.007)
Soil: Bulk density (200 cm depth) (sgbldfie)	0.100 (0.038, 0.163)
Soil: Clay content mass fraction (200 cm depth) (sgclyppt)	-0.078 (-0.157, 0.002)
Soil: Coarse fragments (200 cm depth) (sgcrfvol)	0.077 (0.008, 0.146)
Slope (slope)	0.008 (-0.021, 0.038)
Tasseled cap brightness (tcb_v6)	-0.333 (-0.404, -0.262)
Population (worldpop)	-0.014 (-0.037, 0.010)
River width (river_size)	0.072 (0.039, 0.106)



514 Supplementary Figure 10. Estimated random effects for environmental suitability.

Random effect estimates (mean and 95% UI) are shown for the environmental suitability layer, fit within INLA
using a second-order random walk (RW2) model with standard-normalised suitability values (x-axis) grouped into
bins by quantile; y-axis values indicate effects in logit space.

518

519 5.3.9 Loa loa endemicity

520 The co-occurrence of O. volvulus and the filarial nematode Loa loa complicates onchocerciasis control in some parts 521 of central Africa, (86,87) due to potentially severe complications from ivermectin treatment in individuals with high 522 L. loa microfilariae loads.(88,89) Previous modelling studies have estimated the scale of co-infections between O. 523 volvulus and L. loa and have suggested a substantial burden of co-infection or populations at risk.(90,91) We 524 complemented these studies by calculating the mean number of onchocerciasis cases estimated by our geospatial 525 model in 2018, in areas considered meso- or hyper-endemic for loiasis according to endemicity classifications from 526 ESPEN. We obtained data from ESPEN for loiasis endemicity in 2015 at the level of Implementation Units and 527 combined this with the WorldPop population raster and our onchocerciasis mean prevalence estimates, deriving a 528 mean estimate of 7 146 618 onchocerciasis cases in loiasis-endemic areas (excluding hypo-endemic regions) in 529 2018.

530

531 5.4 Model validation

532 5.4.1 Metrics of predictive validity

- 533 In order to assess the predictive validity of our estimates, we validated our models using spatially stratified five-fold
- 534 out-of-sample cross-validation. To construct each spatial fold, we used a modified bi-tree algorithm to spatially
- aggregate datapoints. This algorithm recursively partitions two-dimensional space, alternating between horizontal

- 536 and vertical splits on the weighted data sample size medians, until the data contained within each spatial partition are
- 537 of a similar sample size. The depth of recursive partitioning is constrained by the target sample size within a
- 538 partition and the minimum number of clusters or pseudo-clusters allowed within each spatial partition (in this case, a
- 539 minimum sample size of 500 was used). These spatial partitions are then allocated to one of five folds for crossvalidation. Temporal partitioning was unstructured (random).
- 540
- 541
- 542 For validation, each geostatistical model was run five times, each time holding out data from one of the folds. A set 543 of out-of-sample predictions were generated by sampling from the posterior predictive distribution for each held-out
- 544 datapoint. A full suite of out-of-sample predictions over the entire dataset was calculated by combining the out-of-
- 545 sample predictions from the five cross-validation runs. Using these out-of-sample predictions, we computed mean
- 546 error (bias), mean absolute error, 95% coverage of the predictive intervals (the proportion of observed out-of-sample
- 547 data that fall within the predicted 95% uncertainty intervals), root-mean squared error (RMSE, which summarises
- 548 error variance), and the correlation of predicted versus observed prevalence at the level of individual datapoints. A
- 549 scatterplot of reported prevalence versus mean out-of-sample predictions is provided in Supplementary Figure 11,
- 550 and validation metrics are summarised in Supplementary Table 9.
- 551
- 552 In addition to performing cross-validation, we also evaluated over-dispersion in the model input data by performing
- 553 posterior predictive checks. Briefly, a binomial count was simulated from each of 1,000 model draws of predictions
- 554 for each input data row. The distribution of predictions provided a good approximation of the observed distribution,
- 555 suggesting that the model is adequately addressing possible over-dispersion in the data.



558 Supplementary Figure 11. Model validation scatterplots for Africa and Yemen.

Reported prevalence versus mean out-of-sample predictions for individual datapoints, by year and country. Vertical
 bars represent 95% UI; red lines indicate equivalence.

561 Supplementary Table 9. Out-of-sample validation metrics at the level of individual datapoints, from five-fold 562 cross-validation

563 Out-of-sample performance was aggregated over 1988–2018 and is also provided for individual model years. Values 564 were computed in prevalence space. N observations: Number of data rows in the full dataset from a given year.

565 Mean abs. error: Mean absolute error. 95% cov.: 95% coverage. Corr.: Correlation. 566

Year	N Observations	Mean error	Mean abs. error	95% cov.	RMSE	Corr.
1988	216	0.158	0.257	0.837	0.298	0.298
1989	310	0.017	0.155	0.934	0.205	0.645
1990	232	-0.081	0.171	0.919	0.209	0.565
1991	58	0.159	0.317	0.639	0.407	-0.003
1992	983	0.040	0.130	0.961	0.174	0.658
1993	866	-0.005	0.175	0.920	0.233	0.554
1994	770	0.060	0.159	0.941	0.210	0.359
1995	58	-0.037	0.124	0.908	0.161	0.500
1996	1 104	0.021	0.093	0.967	0.144	0.709
1997	1 148	-0.008	0.138	0.959	0.189	0.745
1998	1 620	-0.023	0.109	0.970	0.152	0.615
1999	370	-0.052	0.074	0.942	0.126	0.575
2000	958	-0.026	0.089	0.978	0.123	0.814
2001	1 163	-0.010	0.112	0.954	0.163	0.707
2002	885	0.012	0.147	0.913	0.200	0.547
2003	1 082	0.017	0.119	0.953	0.173	0.615
2004	397	0.009	0.113	0.930	0.153	0.658
2005	1 930	0.005	0.144	0.965	0.188	0.533
2006	383	-0.002	0.104	0.727	0.170	0.611
2007	117	-0.017	0.046	0.899	0.083	0.631
2008	367	0.046	0.112	0.902	0.195	0.666
2009	193	-0.010	0.055	0.816	0.092	0.566
2010	133	0.020	0.066	0.904	0.097	0.722
2011	764	0.009	0.097	0.903	0.140	0.620

Year	N Observations	Mean error	Mean abs. error	95% cov.	RMSE	Corr.
2012	543	0.014	0.063	0.934	0.115	0.659
2013	477	-0.014	0.021	0.965	0.046	0.207
2014	397	-0.035	0.051	0.914	0.096	0.344
2015	472	-0.043	0.077	0.853	0.129	0.182
2016	96	0.004	0.033	0.955	0.075	0.138
2017	20	0.062	0.096	0.995	0.189	0.881
2018	4	-0.138	0.138	0.524	0.145	NA
1988–2018	18 116	0.003	0.111	0.924	0.168	0.706

568	6.0 Supplementary references
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- 569
- 570 1. ESPEN. ESPEN. Available from: https://www.espen.org/
- 571 2. WHO | Geographical distribution of onchocerciasis. WHO. World Health Organization;
 572 Available from: https://www.who.int/onchocerciasis/distribution/en/
- 3. Remme JHF. The African programme for onchocerciasis control: preparing to launch.
 Parasitol Today. 1995;11(11):403–6.
- 4. Ngoumou P, Walsh JF, Mace JM. A rapid mapping technique for the prevalence and distribution of onchocerciasis: a Cameroon case study. Ann Trop Med Parasitol.
 1994;88(5):463–74.
- 578 5. Ngoumou P, Walsh JF, Blindness WP for the P of, Diseases UBSP for R and T in T. A
 579 manual for rapid epidemiological mapping of onchocerciasis. 1993; Available from: 580 https://apps.who.int/iris/handle/10665/59537
- 6. Noma M, Nwoke BEB, Nutall I, Tambala PA, Enyong P, Namsenmo A, et al. Rapid
 epidemiological mapping of onchocerciasis (REMO): its application by the African
 Programme for Onchocerciasis Control (APOC). Ann Trop Med Parasitol. 2002;96 Suppl
 1:S29-39.
- 7. Tekle AH, Zouré HGM, Noma M, Boussinesq M, Coffeng LE, Stolk WA, et al. Progress towards onchocerciasis elimination in the participating countries of the African Programme for Onchocerciasis Control: epidemiological evaluation results. Infect Dis Poverty.
 2016;5(1):66.
- 589 8. WHO | African Programme for Onchocerciasis Control: progress report, 2013–2014. Wkly
 590 Epidemiol Rec. 2014;89(49):551–60.

- 591 9. ENVISION. Tanzania Work Plan, FY2018. ENVISION; 2017. Available from:
 592 https://www.ntdenvision.org/sites/default/files/docs/tanzania_work_planfy18.pdf
- 10. de Smet E, Metanmo S, Mbelesso P, Kemata B, Siewe Fodjo JN, Boumédiène F, et al.
 Focus of ongoing onchocerciasis transmission close to Bangui, Central African Republic.
 Pathogens. 2020;9(5):337.
- 596 11. Bof JCM, Maketa V, Bakajika DK, Ntumba F, Mpunga D, Murdoch ME, et al.
 597 Onchocerciasis control in the Democratic Republic of Congo (DRC): challenges in a post598 war environment. Trop Med Int Health. 2015;20(1):48–62.
- 599 12. Makenga Bof JC, Ntumba Tshitoka F, Muteba D, Mansiangi P, Coppieters Y. Review of
 600 the National Program for Onchocerciasis Control in the Democratic Republic of the Congo.
 601 Trop Med Infect Dis. 2019;4(2):92.
- Samuel A, Belay T, Yehalaw D, Taha M, Zemene E, Zeynudin A. Impact of six years
 community directed treatment with ivermectin in the control of onchocerciasis, Western
 Ethiopia. PLOS ONE. 2016;11(3):e0141029.
- 605 14. Gemade EII, Jiya JY, Nwoke BEB, Ogunba EO, Edeghere H, Akoh JI, et al. Human
 606 onchocerciasis: current assessment of the disease burden in Nigeria by rapid
 607 epidemiological mapping. Ann Trop Med Parasitol. 1998;92(sup1):S79–83.
- Republic of Rwanda Ministry of Health. Neglected Tropical Diseases Strategic Plan 20192024. Republic of Rwanda Ministry of Health; 2019. Available from:
 http://rbc.gov.rw/fileadmin/user_upload/guide2019/guide2019/RWANDA%20NTD%20ST
 RATEGIC%20PLAN%202019-2024.pdf
- 612 16. Laggo MCL, Chane F. Onchocerciasis control in South Sudan. South Sudan Med J.
 613 2011;4(3):61-2.
- 614 17. World Bank. Chad, Cameroon Petroleum Development and Pipeline Project:
 615 environmental assessment. Washington, D.C.: World Bank; 1999. Available from:
 616 http://documents.banquemondiale.org/curated/fr/863021468769828252/pdf/multi-page.pdf
- 617 18. Katabarwa MN, Lakwo T, Habomugisha P, Unnasch TR, Garms R, Hudson-Davis L, et al.
 618 After 70 years of fighting an age-old scourge, onchocerciasis in Uganda, the end is in sight.
 619 Int Health. 2018;10(suppl 1):i79–88.
- 620 19. ESPEN Portal. Available from: https://admin.espen.afro.who.int/
- WHO | Global Programme to Eliminate Lymphatic Filariasis. WHO. World Health
 Organization; Available from: http://www.who.int/lymphatic_filariasis/elimination programme/en/
- 624 21. Graham MH. Confronting Multicollinearity in Ecological Multiple Regression. Ecology.
 625 2003;84(11):2809–15.

- 626 22. Faraway JJ. Linear Models with R. Boca Raton: CRC Press; 2005.
- Prost A, Thylefors B, Pairault C. Methods of mass epidemiological evaluation of
 onchocerciasis: their utilization in a vector control programme. Expert Comm Epidemiol
 Onchocerciasis. 1975;20.
- 630 24. O'Hanlon SJ, Slater HC, Cheke RA, Boatin BA, Coffeng LE, Pion SDS, et al. Model-based
 631 geostatistical mapping of the prevalence of Onchocerca volvulus in West Africa. PLoS
 632 Negl Trop Dis. 2016;10(1):e0004328.
- 633 25. World Health Organization. Supplemental Guidelines for Rapid Epidemiological Mapping
 634 of Onchocerciasis (REMO). Geneva, Switzerland: World Health Organization; 1995.
 635 Report No.: Document TDR/TDF/ONCHO/95.1. Available from:
 636 https://www.who.int/tdr/publications/tdr-research-publications/oncho-guidelines/en/
- 26. Zouré HG, Noma M, Tekle AH, Amazigo UV, Diggle PJ, Giorgi E, et al. The geographic
 distribution of onchocerciasis in the 20 participating countries of the African Programme
 for Onchocerciasis Control: (2) pre-control endemicity levels and estimated number
 infected. Parasit Vectors. 2014;7(1):326.
- 641 27. Gallin M, Adams A, Kruppa TF, Gbaguidi EA, Massougbodji A, Sadeler BC, et al.
 642 Epidemiological studies of onchocerciasis in southern Benin. Trop Med Parasitol Off Organ
 643 Dtsch Tropenmedizinische Ges Dtsch Ges Tech Zusammenarbeit GTZ. 1993;44(2):69–74.
- Boussinesq M, Chippaux JP, Ernould JC, Quillevere D, Prod'hon J. Effect of repeated
 treatments with ivermectin on the incidence of onchocerciasis in northern Cameroon. Am J
 Trop Med Hyg. 1995;53(1):63–7.
- 647 29. Mendoza Aldana J, Piechulek H, Maguire J. Forest onchocerciasis in Cameroon: its
 648 distribution and implications for selection of communities for control programmes. Ann
 649 Trop Med Parasitol. 1997;91(1):79–86.
- 30. Katabarwa MN, Eyamba A, Nwane P, Enyong P, Kamgno J, Kueté T, et al. Fifteen years of
 annual mass treatment of onchocerciasis with ivermectin have not interrupted transmission
 in the west region of Cameroon. J Parasitol Res. 2013;2013. Available from:
 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3652197/
- 654 31. Kamgno J, Boussinesq M. Hyperendemic loaiasis in the Tikar plain, shrub savanna region
 655 of Cameroon. Bull Soc Pathol Exot 1990. 2001;94(4):342–6.
- Katabarwa MN, Eyamba A, Chouaibou M, Enyong P, Kuété T, Yaya S, et al. Does
 onchocerciasis transmission take place in hypoendemic areas? a study from the North
 Region of Cameroon. Trop Med Int Health. 2010;15(5):645–52.
- 33. Katabarwa MN, Eyamba A, Nwane P, Enyong P, Yaya S, Baldiagaï J, et al. Seventeen
 years of annual distribution of ivermectin has not interrupted onchocerciasis transmission in
 North Region, Cameroon. Am J Trop Med Hyg. 2011;85(6):1041–9.

662 34. Kamga HLF, Shey DN, Assob JCN, Njunda AL, Nde Fon P, Njem PK. Prevalence of 663 onchocerciasis in the Fundong Health District, Cameroon after 6 years of continuous 664 community-directed treatment with ivermectin. Pan Afr Med J. 2011;10. Available from: 665 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3240934/ 666 35. Wanji S, Kengne-Ouafo JA, Esum ME, Chounna PWN, Adzemye BF, Eyong JEE, et al. Relationship between oral declaration on adherence to ivermectin treatment and 667 668 parasitological indicators of onchocerciasis in an area of persistent transmission despite a 669 decade of mass drug administration in Cameroon. Parasit Vectors. 2015;8. Available from: 670 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4696282/ 671 36. Kamga GR, Dissak-Delon FN, Nana-Djeunga HC, Biholong BD, Ghogomu SM, Souopgui J, et al. Important progress towards elimination of onchocerciasis in the West Region of 672 673 Cameroon. Parasit Vectors. 2017;10. Available from: 674 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5543544/ 675 37. Kamga GR, Dissak-Delon FN, Nana-Djeunga HC, Biholong BD, Mbigha-Ghogomu S, 676 Souopgui J, et al. Still mesoendemic onchocerciasis in two Cameroonian community-677 directed treatment with ivermectin projects despite more than 15 years of mass treatment. Parasit Vectors. 2016;9(1):581. 678 679 38. Bakajika D, Senyonjo L, Enyong P, Oye J, Biholong B, Elhassan E, et al. On-going 680 transmission of human onchocerciasis in the Massangam health district in the West Region 681 of Cameroon: Better understanding transmission dynamics to inform changes in 682 programmatic interventions. PLoS Negl Trop Dis. 2018;12(11):e0006904. 683 39. Katabarwa M, Eyamba A, Habomugisha P, Lakwo T, Ekobo S, Kamgno J, et al. After a 684 decade of annual dose mass ivermectin treatment in Cameroon and Uganda, onchocerciasis 685 transmission continues. Trop Med Int Health TM IH. 2008;13(9):1196-203. 686 40. Kennedy MH, Bertocchi I, Hopkins AD, Meredith SE. The effect of 5 years of annual 687 treatment with ivermectin (Mectizan) on the prevalence and morbidity of onchocerciasis in the village of Gami in the Central African Republic. Ann Trop Med Parasitol. 688 689 2002;96(3):297-307. 690 41. Taye A, Gebre-Michael T, Taticheff S. Onchocerciasis in Gilgel Ghibe River Valley 691 southwest Ethiopia. East Afr Med J. 2000;77(2):116-20. 692 42. Mengistu G, Balcha F, Britton S. Co-infection of Onchocerca volvulus and intestinal 693 helminths in indigenous and migrant farmers in southwest Ethiopia. Ethiop Med J. 694 2002;40(1):19-27. 695 43. Legesse M, Balcha F, Erko B. Status of onchocerciasis in Teppi area, Southwestern Ethiopia, after four years of annual community-directed treatment with ivermectin. Ethiop J 696 697 Health Dev. 2010;24(1). Available from: https://www.ajol.info/index.php/ejhd/article/view/62945 698

699 44. Dori GU, Belay T, Belete H, Panicker KN, Hailu A. Parasitological and clinico-700 epidemiological features of onchocerciasis in West Wellega, Ethiopia. J Parasit Dis Off 701 Organ Indian Soc Parasitol. 2012;36(1):10-8. 702 45. Dana D, Debalke S, Mekonnen Z, Kassahun W, Suleman S, Getahun K, et al. A 703 community-based cross-sectional study of the epidemiology of onchocerciasis in unmapped 704 villages for community directed treatment with ivermectin in Jimma Zone, southwestern 705 Ethiopia. BMC Public Health. 2015;15:595. 706 46. Fobi G, Mourou Mbina JR, Ozoh G, Kombila M, Agaya C, Olinga Olinga JM, et al. 707 Onchocerciasis in the area of Lastourville, Gabon. Clinical and entomological aspects. Bull 708 Soc Pathol Exot 1990. 2006;99(4):269-71. 709 47. Anosike JC, Onwuliri CO. Studies on filariasis in Bauchi State, Nigeria. II. The prevalence 710 of human filariasis in Darazo Local Government area. Appl Parasitol. 1994;35(4):242-50. 711 48. Osue HO, Inabo HI, Yakubu SE, Audu PA, Galadima M, Odama LE, et al. Impact of 712 eighteen-year varied compliance to onchocerciasis treatment with ivermectin in sentinel 713 savannah agrarian communities in Kaduna State of Nigeria. ISRN Parasitol. 714 2013;2013:960168. 715 49. Anosike JC, Celestine null, Onwuliri OE, Onwuliri VA. The prevalence, intensity and 716 clinical manifestations of Onchocerca volvulus infection in Toro local government area of 717 Bauchi State, Nigeria. Int J Hyg Environ Health. 2001;203(5-6):459-64. 718 50. Umeh RE. The causes and profile of visual loss in an onchocerciasis-endemic forest-719 savanna zone in Nigeria. Ophthalmic Epidemiol. 1999;6(4):303-15. 720 51. Nmorsi OPG, Oladokun I a. A, Egwunyenga OA, Oseha E. Eye lesions and onchocerciasis 721 in a rural farm settlement in Delta state, Nigeria. Southeast Asian J Trop Med Public 722 Health. 2002;33(1):28–32. 723 52. Okoye IC, Onwuliri CO. Epidemiology and psycho-social aspects of onchocercal skin diseases in northeastern Nigeria. Filaria J. 2007;6:15. 724 725 53. Rebecca SN, Akinboye DO, Abdulazeez AA. Onchocerciasis and plasmodiasis: concurrent 726 infection in Garaha-Dutse community, Adamawa State Nigeria. Biomed Res. 2008;19(2). 727 Available from: https://www.alliedacademies.org/abstract/onchocerciasis-and-728 plasmodiasis-concurrent-infectionrnin-garahadutse-community-adamawa-state-nigeria-729 802.html 730 54. Emukah E, Nimzing J, Ezeabikwa M, Obiezu J, Okpala N, Miri E, et al. Impact assessment 731 of repeated annual ivermectin on ocular and clinical onchocerciasis 14 years of annual mass 732 drug administration in eight sentinel villages, southeast Nigeria. Am J Trop Med Hyg. 733 2010;83(Suppl 5):22. 734 55. Akinbo FO, Okaka CE. Hyperendemicity of onchocerciasis in Ovia Northeast Local Government Area, Edo State, Nigeria. East Afr J Public Health. 2010;7(1):84-6. 735

736 737 738 739	56.	Akinboye, D.O, Okwong, E, N A, Agbolade FO, et al. Onchocerciasis among inhabitants of Ibarapa local government community of Oyo state, Nigeria. Biomed Res. 2010;21(2). Available from: https://www.alliedacademies.org/abstract/onchocerciasis-among- inhabitants-of-ibarapa-local-governmentrncommunity-of-oyo-state-nigeria-1287.html
740 741	57.	Uttah E, Ibeh DC. Multiple filarial species microfilaraemia: a comparative study of areas with endemic and sporadic onchocerciasis. J Vector Borne Dis. 2011;48(4):197–204.
742 743 744 745 746	58.	Evans DS, Alphonsus K, Umaru J, Eigege A, Miri E, Mafuyai H, et al. Status of onchocerciasis transmission after more than a decade of mass drug administration for onchocerciasis and lymphatic filariasis elimination in Central Nigeria: challenges in coordinating the Stop MDA Decision. PLoS Negl Trop Dis. 2014;8(9). Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4169246/
747 748	59.	Eyo J, Onyishi G, Ugokwe C. Rapid epidemiological assessment of onchocerciasis in a tropical semi-urban community, enugu state, Nigeria. Iran J Parasitol. 2013;8(1):145–51.
749 750 751	60.	Koroma JB, Sesay S, Conteh A, Koudou B, Paye J, Bah M, et al. Impact of five annual rounds of mass drug administration with ivermectin on onchocerciasis in Sierra Leone. Infect Dis Poverty. 2018;7(1):30.
752 753	61.	Dogba PK, Komi BT. The evaluation of endemic onchocerciasis in the Amou prefecture (Togo). Bull Soc Pathol Exot 1990. 1994;87(2):110–1.
754 755 756	62.	Golden A, Faulx D, Kalnoky M, Stevens E, Yokobe L, Peck R, et al. Analysis of age- dependent trends in Ov16 IgG4 seroprevalence to onchocerciasis. Parasit Vectors. 2016;9(1):338.
757 758 759 760 761	63.	Komlan K, Vossberg PS, Gantin RG, Solim T, Korbmacher F, Banla M, et al. Onchocerca volvulus infection and serological prevalence, ocular onchocerciasis and parasite transmission in northern and central Togo after decades of Simulium damnosum s.l. vector control and mass drug administration of ivermectin. PLoS Negl Trop Dis. 2018;12(3):e0006312.
762 763 764 765	64.	Katabarwa MN, Lakwo T, Habomugisha P, Agunyo S, Byamukama E, Oguttu D, et al. Transmission of Onchocerca volvulus continues in Nyagak-Bondo focus of northwestern Uganda after 18 years of a single dose of annual treatment with ivermectin. Am J Trop Med Hyg. 2013;89(2):293–300.
766 767 768 769	65.	Luroni LT, Gabriel M, Tukahebwa E, Onapa AW, Tinkitina B, Tukesiga E, et al. The interruption of Onchocerca volvulus and Wuchereria bancrofti transmission by integrated chemotherapy in the Obongi focus, North Western Uganda. PloS One. 2017;12(12):e0189306.
770 771 772 773	66.	Murray CJL, Callender CSKH, Kulikoff XR, Srinivasan V, Abate D, Abate KH, et al. Population and fertility by age and sex for 195 countries and territories, 1950–2017: a systematic analysis for the Global Burden of Disease Study 2017. The Lancet. 2018;392(10159):1995–2051.

774 67. Ramsay JO, Wickham H, Graves S, Hooker G. fda: Functional Data Analysis. 2018. 775 Available from: https://CRAN.R-project.org/package=fda 776 68. Byrd RH, Lu P, Nocedal J, Zhu C. A limited memory algorithm for bound constrained 777 optimization. SIAM J Sci Comput. 1995;16(5):1190-208. 778 69. R: The R Project for Statistical Computing. Available from: https://www.r-project.org/ 779 70. Kirkwood B, Smith P, Marshall T, Prost A. Variations in the prevalence and intensity of 780 microfilarial infections by age, sex, place and time in the area of the Onchocerciasis Control 781 Programme. Trans R Soc Trop Med Hyg. 1983;77(6):857-61. 782 71. Vivas-Martínez S, Basáñez MG, Botto C, Rojas S, García M, Pacheco M, et al. Amazonian 783 onchocerciasis: parasitological profiles by host-age, sex, and endemicity in southern 784 Venezuela. Parasitology. 2000;121(5):513-25. 785 72. Anderson RM, Basáñez MG, Boussinesq M. Population biology of human onchocerciasis. 786 Philos Trans R Soc Lond B Biol Sci. 1999;354(1384):809-26. 787 73. Coffeng LE, Pion SDS, O'Hanlon S, Cousens S, Abiose AO, Fischer PU, et al. 788 Onchocerciasis: the pre-control association between prevalence of palpable nodules and 789 skin microfilariae. PLoS Negl Trop Dis. 2013;7(4):e2168. 790 74. Hamley JID, Milton P, Walker M, Basáñez MG. Modelling exposure heterogeneity and 791 density dependence in onchocerciasis using a novel individual-based transmission model, 792 EPIONCHO-IBM: Implications for elimination and data needs. PLoS Negl Trop Dis. 793 2019;13(12):e0007557. 794 75. Gebrezgabiher G, Mekonnen Z, Yewhalaw D, Hailu A. Reaching the last mile: main 795 challenges relating to and recommendations to accelerate onchocerciasis elimination in 796 Africa. Infect Dis Poverty. 2019;8(1):60. 797 76. Vlaminck J, Fischer PU, Weil GJ. Diagnostic tools for onchocerciasis elimination 798 programs. Trends Parasitol. 2015;31(11):571-82. 799 77. Turner HC, Walker M, Churcher TS, Basáñez MG. Modelling the impact of ivermectin on 800 River Blindness and its burden of morbidity and mortality in African Savannah: EpiOncho 801 projections. Parasit Vectors. 2014;7(1):241. 802 78. Remme. The global burden of onchocerciasis in 1990. Geneva, Switzerland: World Health 803 Organization; 2004 p. 26. Available from: 804 https://www.who.int/apoc/publications/bod onchocerciasis1990 the global burden of on chocerciasis in 1990.pdf 805 806 79. Unnasch TR, Golden A, Cama V, Cantey PT. Diagnostics for onchocerciasis in the era of 807 elimination. Int Health. 2018;10(suppl 1):i20-6.

- 808 80. Golding N, Burstein R, Longbottom J, Browne AJ, Fullman N, Osgood-Zimmerman A, et
 809 al. Mapping under-5 and neonatal mortality in Africa, 2000–15: a baseline analysis for the
 810 Sustainable Development Goals. The Lancet. 2017;390(10108):2171–82.
- 811 81. Cromwell EA, Osborne JCP, Unnasch TR, Basáñez MG, Gass KM, Barbre KA, et al.
 812 Predicting the environmental suitability for onchocerciasis in Africa as an aid to elimination 813 planning. PLoS Negl Trop Dis. 2021;15(7):e0008824.
- 814 82. Gómez-Rubio V, Palmí-Perales F. Multivariate posterior inference for spatial models with
 815 the integrated nested Laplace approximation. J R Stat Soc Ser C Appl Stat.
 816 2019;68(1):199–215.
- 83. Lindgren F, Rue H, Lindström J. An explicit link between Gaussian fields and Gaussian
 Markov random fields: the stochastic partial differential equation approach. Stat Methodol
 Ser B. 2011;73(4):423–98.
- 84. Blangiardo M, Cameletti M, Baio G, Rue H. Spatial and spatio-temporal models with RINLA. Spat Spatio-Temporal Epidemiol. 2013;7:39–55.
- 822 85. Lindgren F, Rue H. Bayesian Spatial Modelling with R- INLA. J Stat Softw.
 823 2015;63(19):1–25.
- 86. Wanji S, Ndongmo WPC, Fombad FF, Kengne-Ouafo JA, Njouendou AJ, Tchounkeu YFL,
 et al. Impact of repeated annual community directed treatment with ivermectin on loiasis
 parasitological indicators in Cameroon: Implications for onchocerciasis and lymphatic
 filariasis elimination in areas co-endemic with Loa loa in Africa. PLoS Negl Trop Dis.
 2018;12(9):e0006750.
- 87. Zouré HGM, Wanji S, Noma M, Amazigo UV, Diggle PJ, Tekle AH, et al. The geographic
 distribution of Loa loa in Africa: results of large-scale implementation of the rapid
 assessment procedure for loiasis (RAPLOA). PLoS Negl Trop Dis. 2011;5(6):e1210.
- 832 88. Gardon J, Gardon-Wendel N, Demanga-Ngangue, Kamgno J, Chippaux JP, Boussinesq M.
 833 Serious reactions after mass treatment of onchocerciasis with ivermectin in an area endemic
 834 for Loa loa infection. The Lancet. 1997;350(9070):18–22.
- 835 89. Boussinesq M, Gardon J, Gardon-Wendel N, Chippaux JP. Clinical picture, epidemiology
 836 and outcome of Loa-associated serious adverse events related to mass ivermectin treatment
 837 of onchocerciasis in Cameroon. Filaria J. 2003;2(1):S4.
- 838 90. Vinkeles Melchers NVS, Coffeng LE, Boussinesq M, Pedrique B, Pion SDS, Tekle AH, et
 839 al. Projected number of people with onchocerciasis–loiasis coinfection in Africa, 1995 to
 840 2025. Clin Infect Dis. 2020;70(11):2281–9.
- 841 91. Cano J, Basáñez MG, O'Hanlon SJ, Tekle AH, Wanji S, Zouré HG, et al. Identifying co842 endemic areas for major filarial infections in sub-Saharan Africa: seeking synergies and
 843 preventing severe adverse events during mass drug administration campaigns. Parasit
 844 Vectors. 2018;11(1):70.